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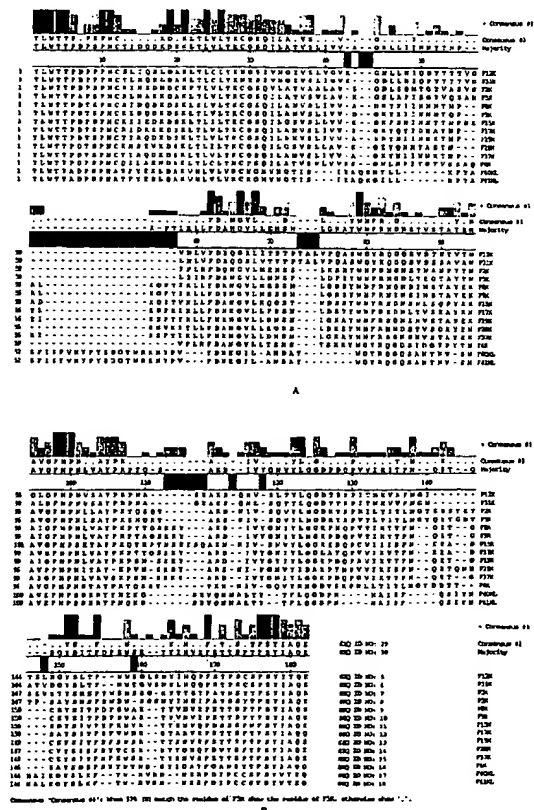
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(54) Title: ALTERNATIVELY TARGETED ADENOVIRUS

## (57) Abstract

The present invention provides a recombinant protein having an amino terminus of an adenoviral fiber protein and having a trimerization domain. A fiber incorporating such a protein exhibits reduced affinity for a native substrate than does a wild-type adenoviral fiber trimer. The present invention further provides an adenovirus incorporating the recombinant protein of the present invention.



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## ALTERNATIVELY TARGETED ADENOVIRUS

### TECHNICAL FIELD OF THE INVENTION

The present invention relates to an alternately targeted adenovirus and  
5 includes methods for producing and purifying such viruses as well as protein  
modifications mediating alternate targeting.

### BACKGROUND OF THE INVENTION

The various physiological responses of a host animal to the presence of a  
10 virus depend on the different ways such viruses interact with the host animal, each  
of which is first mediated by the surface of the virus ("the virion"). The  
adenoviral virion is a non-enveloped icosahedron about 65-80 nm in diameter  
(Horne *et al.*, *J. Mol. Biol.*, 1, 84-86 (1959)). It comprises 252 capsomeres -- 240  
hexons and 12 pentons (Ginsberg *et al.*, *Virology*, 28, 782-83 (1966)) -- derived  
15 from three viral proteins (proteins II, III, and IV) (Maizel *et al.*, *Virology*, 36, 115-  
25 (1968); Weber *et al.*, *Virology*, 76, 709-24 (1977)). Proteins IX, VI, and IIIa,  
also present, stabilize the virion (Stewart *et al.*, *Cell*, 67, 145-54 (1991); Stewart *et al.*,  
*EMBO J.*, 12(7), 2589-99 (1993)).

The hexon provides structure and form to the capsid (Pettersson, in *The*  
20 *Adenoviruses*, pp. 205-270, Ginsberg, ed., (Plenum Press, New York, NY, 1984)),  
and is a homotrimer of the protein II (Roberts *et al.*, *Science*, 232, 1148-1151  
(1986)). The hexon provides the main antigenic determinants of the virus, and it  
also contributes to the serotype specificity of the virus (Watson *et al.*, *J. Gen.*  
*Virol.*, 69, 525-35 (1988); Wolfort *et al.*, *J. Virol.*, 62, 2321-28 (1988); Wolfort *et al.*,  
25 *J. Virol.*, 56, 896-903 (1985); Crawford-Miksza *et al.*, *J. Virol.*, 70, 1836-44  
(1996)).

The hexon trimer is comprised of a pseudohexagonal base and a triangular  
top formed of three towers (Roberts *et al.*, *supra*; Athappilly *et al.*, *J. Mol. Biol.*,  
242, 430-455 (1994)). The base pedestal consists of two tightly packed eight-  
30 stranded antiparallel beta barrels stabilized by an internal loop. The predominant  
antigenic and serotype-specific regions of the hexon appear to be in loops 1 and 2  
(i.e., LI or I1, and LII or I2, respectively), within which are seven discrete  
hypervariable regions (HVR1 to HVR7) varying in length and sequence between  
adenoviral serotypes (Crawford-Miksza *et al.*, *supra*).

35 The penton contains a base, which is bound to the capsid, and a fiber,  
which is non-covalently bound to and projects from the penton base. The penton  
base, consisting of protein III, is highly conserved among serotypes of adenovirus.

and (except for the enteric adenovirus Ad40 and Ad41) it has five RGD tripeptide motifs (Neumann *et al.*, *Gene*, 69, 153-57 (1988)). These RGD tripeptides apparently mediate adenoviral binding to  $\alpha_v$  integrins, a family of a heterodimeric cell-surface receptors that also interact with the extracellular matrix and play important roles in cell signaling (Hynes, *Cell*, 69, 11-25 (1992)). These RGD tripeptides also play a role in endocytosis of the virion (Wickham *et al.* (1993), *supra*; Bai *et al.*, *J. Virol.*, 67, 5198-3205 (1993)).

The adenoviral fiber is a homotrimer of the adenoviral polypeptide IV (Devaux *et al.*, *J. Molec. Biol.*, 215, 567-88 (1990)), which has three discrete domains. The amino-terminal "tail" domain attaches non-covalently to the penton base. A relatively long "shaft" domain, comprising a variable number of repeating 15 residue  $\beta$ -sheets motifs, extends outwardly from the vertices of the viral particle (Yeh *et al.*, *Virus Res.*, 33, 179-98 (1991)). Lastly, about 200 residues at the carboxy-terminus form the "knob" domain. Functionally, the knob mediates both primary viral binding to cellular proteins and fiber trimerization (Henry *et al.*, *J. Virol.*, 68(8), 5239-46 (1994)). Trimerization also appears necessary for the amino terminus of the fiber to properly associate with the penton base (Novelli *et al.*, *Virology*, 185, 365-76 (1991)). In addition to recognizing cell receptors and binding the penton base, the fiber contributes to serotype integrity and mediates nuclear localization. Moreover, adenoviral fibers from several serotypes are glycosylated (see, e.g., Mullis *et al.*, *J. Virol.*, 64(11), 5317-23 (1990); Hong *et al.*, *J. Virol.*, 70(10), 7071-78 (1996); Chroboczek *et al.*, *Adenovirus Fiber*, p. 163-200 in "The Molecular Repertoire of Adenoviruses I. Virion Structure and Function," W. Doerfler and P. Böhm, eds. (Springer, NY 1995)).

Fiber proteins from different adenoviral serotypes differ considerably. For example, the number of shaft repeats differs between adenoviral serotypes (Green *et al.*, *EMBO J.*, 2, 1357-65 (1983)). Moreover, the knob regions from the closely related Ad2 and Ad5 serotypes are only 63% similar at the amino acid level (Chroboczek *et al.*, *Virology*, 186, 280-85 (1992)), and Ad2 and Ad3 fiber knobs are only 20% identical (Signas *et al.*, *J. Virol.*, 53, 672-78 (1985)). In contrast, the penton base sequences of Ad5 and Ad2 are 99% identical. Despite these differences in the knob region, a sequence comparison of even the Ad2 and Ad3 fiber genes demonstrates distinct regions of conservation, most of which are also conserved among the other human adenoviral fibers (see, e.g., Figures 1A-2B)

One interaction between the adenoviral virion and the host animal is the process of cellular infection, during which the wild-type virion first binds the cell surface by means of a cellular adenoviral receptor (AR) (e.g., the coxsackievirus

and adenovirus receptor (CAR), the MHC class I receptor, etc. (Bergelson *et al.*, *Science*, 275, 1320-23 (1997); Tanako *et al.*, *Proc. Nat. Acad. Sci. (USA)*, 94, 3352-56 (1997)). Hong *et al.*, *EMBO J.*, 16(9), 2294-06 (1997)). After attachment to an AR, the virus binds  $\alpha_v$  integrins. Following attachment to these cell surface proteins, infection proceeds by receptor-mediated internalization of the virus into endocytotic vesicles (Svensson *et al.*, *J. Virol.*, 51, 687-94 (1984); Chardonnet *et al.*, *Virology*, 40, 462-77 (1970)). Within the cell, virions are disassembled (Greber *et al.*, *Cell*, 75, 477-86 (1993)), the endosome disrupted (Fitzgerald *et al.*, *Cell*, 32, 607-17 (1983)), and the viral particles transported to the nucleus via the nuclear pore complex (Dales *et al.*, *Virology*, 56, 465-83 (1973)). As most adenoviral serotypes interact with cells through broadly disseminated cell surface proteins, the natural range of host cells infected by adenovirus is broad.

In addition to cellular infection, host animals react defensively to the presence of adenoviral virions through mechanisms that reduce the effective free titer of the virus. For example, host immune systems, upon exposure to a given adenoviral serotype, can efficiently develop neutralizing antibodies, greatly reducing the effective free titer of the virus upon repeat administration (see, e.g., Setoguchi *et al.*, *Am. J. Respir. Cell. Mol. Biol.*, 10, 369-77 (1994); Kass-Eisler *et al.*, *Gene Ther.*, 1, 395-402 (1994); Kass-Eisler *et al.*, *Gene Ther.*, 3, 154-62 (1996)). Interestingly, such antibodies typically are directed against the same determinants of adenoviral serotype specificity, and are primarily directed to the hypervariable hexon regions and, to some extent, fiber and penton base domains (Watson *et al.*, *supra*; Wolfort *et al.* (1988), *supra*; Wolfort *et al.* (1985), *supra*; Crawford-Miksza *et al.*, *supra*). Of course, the presence of adenoviruses agglutinates red blood cells in humans in a serotype-dependent manner (Hierholzer, *J. Infect. Diseases*, 123(4), 541-50 (1973)). Additionally, adenoviral virions are actively scavenged from the circulation by cells of the reticuloendothelial system (RES) (see, e.g., Worgall *et al.*, *Hum Gene Ther.*, 8, 1675-84 (1997); Wolff *et al.*, *J. Virol.*, 71(1), 624-29 (1997)). In such a response, Kupffer cells, endothelial liver cells, or other RES cells scavenge the virus from the circulation (see generally, Moghini *et al.*, *Crit. Rev. Ther. Drug Carrier Sys.*, 11(1), 31-59 (1994); Van Rooijen *et al.*, *J. Leuk. Biol.*, 62, 702-09 (1997)). For example, virions can become opsonized, possibly through interaction between collectins and glycosylated viral proteins, triggering recognition by such RES cells; alternatively, such cells may recognize charged amino acid residues on the virion surface (see Hansen *et al.*, *Immunobiol.*, 199(2), 165-89 (1998); Jahrling *et al.*, *J. Med. Virol.*, 12(1), 1-16 (1983)).

Based on the popularity of adenoviruses as gene transfer vectors, efforts have been made to increase the ability of adenovirus to enter certain cells, e.g., those few cells it does not infect, an approach referred to as "targeting" (see, e.g., International Patent Application WO 95/26412 (Curiel *et al.*), International Patent Application WO 94/10323 (Spooner *et al.*), U.S. Patent 5,543,328 (McClelland *et al.*), International Patent Application WO 94/24299 (Cotten *et al.*)). Of course, while the ability to target adenoviruses to certain cell types is an important goal, far more desirable is an adenovirus which infects only a desired cell type, an approach referred to as "alternative targeting." However, to exclusively target a virus, its native affinity for host cell ARs must first be abrogated, producing a recombinant adenovirus incapable of productively infecting the full set of natural adenoviral target cells. Efforts aimed at abrogating native adenoviral cell affinity have focused logically on changing the fiber knob. These efforts have proven disappointing, largely because they fail to preserve the important fiber protein functions of stable trimerization and penton base binding (Spooner *et al.*, *supra*). Moreover, replacement of the fiber knob with a cell-surface ligand (McClelland *et al.*, *supra*) produces a virus only suitable for infecting a cell type having that ligand. Such a strategy produces a virus having many of the same targeting problems associated with wild-type adenoviruses (in which fiber trimerization and cellular tropism are mediated by the same protein domain), thus decreasing the flexibility of the vector. Moreover, due to the necessity of having a propagating cell line, and the integral connection between the fiber trimerization and targeting functions, obtaining a mutant virus with substituted targeting is difficult. For example, removing the fiber knob and replacing it with a non-trimerizing ligand (e.g., Spooner *et al.*, McClelland *et al.*, *supra*) results in a virus lacking appreciable fiber protein.

Aside from the broad natural tropism of the virus noted above, the non-infectious interactions between adenovirus and the host also pose problems for using adenovirus as gene transfer vectors. Such interactions effectively reduce the free titer of a given dose of adenovirus beneath that which is clinically effective. As such, there is currently a need for an adenovirus exhibiting reduced affinity for such natural interactions with a host animal (e.g., target cell affinity, innate or acquired immune surveillance, etc). Moreover, there is a need for such a virus which is able to deliver and express a desired transgene within a predefined tissue – an alternatively targeted virus.

### BRIEF SUMMARY OF THE INVENTION

The present invention provides a recombinant protein having an amino terminus of an adenoviral fiber protein and having a trimerization domain. A fiber incorporating such a protein exhibits reduced affinity for a native substrate than  
5 does a wild-type adenoviral fiber trimer. The present invention further provides an adenovirus incorporating the recombinant protein of the present invention.

The present invention is useful in a variety of gene-transfer applications, *in vitro* and *in vivo*, as a vector for delivering a desired gene to a cell with minimal ectopic infection. Specifically, the present invention permits more efficient  
10 production and construction of safer vectors for gene transfer applications. The present invention is also useful as a research tool by providing methods and reagents for the study of adenoviral attachment and infection of cells and in a method of assaying receptor-ligand interaction. Similarly, the recombinant fiber protein can be used in receptor-ligand assays and as adhesion proteins *in vitro* or *in vivo*.  
15 Additionally, the present invention provides reagents and methods permitting biologists to investigate the cell biology of viral growth and infection. Thus, the vectors of the present invention are highly useful in biological research.

### DESCRIPTION OF THE DRAWINGS

20 Figures 1A and 1B sets forth a comparison of the amino acid sequences of the non-group B serotype fiber knobs (SEQ ID NOs: 5-18) using the Clustal method with PAM100 residue weight table. The height of the bars at the top of each row of sequence comparison correlates to the degree of homology. Consensus and majority sequences are indicated as SEQ ID NOs: 29 and 30,  
25 respectively.

Figures 2A and 2B sets forth a comparison of the amino acid sequences of the group B serotype fiber knobs (SEQ ID NOs: 19-25) using the J. Hein method with PAM250 residue weight. The height of the bars at the top of each row of sequence comparison correlates to the degree of homology. Consensus and  
30 majority sequences are indicated as SEQ ID NOs: 31 and 32, respectively.

### DETAILED DESCRIPTION OF THE INVENTION

#### Recombinant Protein

The present invention provides a recombinant adenoviral fiber protein  
35 having an amino terminus derived from an adenoviral fiber protein and having a trimerization domain. A trimer including such a recombinant protein exhibits reduced affinity for a native substrate, such as an antibody, collectins, opsins, a cellular binding site, etc. (i.e., native to the serotype from which the shaft, and

particularly the amino-terminus, is drawn) as compared to a native adenoviral fiber trimer. The trimer can be a homotrimer or a heterotrimer of different fiber monomers. Any modification of the monomeric units reducing the affinity of the resulting trimer for its native cell surface binding site (i.e., a native AR) is within  
5 the scope of the invention. Preferably, the reduction in affinity is a substantial reduction in affinity (such as at least an order of magnitude, and preferably more) relative to the unmodified corresponding fiber.

As mentioned, where a trimerization domain is itself a ligand for a native cell surface binding site, fiber proteins possessing such trimerization domains  
10 present some of the same problems for targeting as native adenoviral fiber trimerization domains. Therefore, the trimerization domain of the inventive protein invention preferably is not a ligand for the CAR or MHC-I cell surface proteins. Most preferably, the non-native trimerization domain is not a ligand for any native adenoviral cell-surface binding site, whether the site is an AR or other  
15 cell surface binding site. As is discussed herein, adenoviruses incorporating such proteins exhibit reduced ability to appreciably infect cells via native AR proteins, and can serve as efficient source vectors for engineering alternatively targeted vectors. Therefore, while the trimerization domain preferably is not a ligand for a cell surface binding site, the entire trimer can be such a ligand (e.g., by virtue of a  
20 non-native ligand as discussed herein). Moreover, the trimerization domain can be a ligand for a substrate other than a native cell surface binding site, as such trimerization-ligands do not present the same concern for cell targeting as do trimerization domains which are ligands for cell surface binding sites. Thus, for example, the non-native trimerization domain can be a ligand for a substrate on an  
25 affinity column, on a blood-borne molecule, or even on a cell surface when it is not a native cell-surface binding site (e.g., on a cell engineered to express a substrate cell surface protein not native to the unmodified cell type).

The recombinant fiber protein can lack a sizable number of residues, or even identifiable domains, as herein described. For example, the protein can lack  
30 the native knob domain; it can lack one or more native shaft  $\beta$ -sheet repeats, or it can be otherwise truncated. Thus, a recombinant fiber protein can have any desired modification so long as it trimerizes when produced by a eukaryotic cell. Furthermore, a recombinant fiber protein preferably is not modified appreciably at the amino terminus (e.g., the amino-terminus of a monomer preferably consists  
35 essentially of the native fiber amino-terminus) to ensure that a fiber incorporating the recombinant fiber protein interacts properly with the penton base. Hence, the present invention also provides a composition of matter comprising a recombinant



fiber protein of the present invention and an adenoviral penton base. Preferably, the recombinant fiber protein and the penton base associate much in the same manner as wild-type fibers and penton bases. Of course, the penton base can also be modified, for example, to include a non-native ligand, for example as is  
5 described in U.S. Patent 5,559,099 (Wickham *et al.*).

In one embodiment, the fiber is modified to render it less able to interact with the innate or acquired host immune system. For example, one or more amino acids of the native fiber protein can be mutated to render the recombinant fiber protein less able to be recognized by neutralizing antibodies than a wild-type fiber  
10 (see, e.g., International Patent Application WO 98/40509 (Crystal *et al.*). The fiber also can be modified to lack one or more amino acids mediating interaction with the RES. For example, the fiber can be mutated to lack one or more glycosylation or phosphorylation sites, or the fiber (or virus containing the fiber) can be produced in the presence of inhibitors of glycosylation or phosphorylation.  
15 Similarly, the fiber (or other protein within the virus) can be conjugated to a lipid derivative of polyethylene glycol (PEG) comprising a primary amine group, an epoxy group, or a diacylglycerol group (see, e.g., Kilbanov *et al.*, *FEBS Lett.*, 268, 235 (1990); Senior *et al.*, *Biochem. Biophys. Acta.*, 1062, 11 (1991); Allen *et al.*, *Biochem. Biophys. Acta.*, 1066, 29 (1991); Mori *et al.*, *FEBS Lett.*, 284, 263  
20 (1991)) to avoid collectin and/or opsonin binding or scavenging by Kupffer (or other RES) cells.

A recombinant fiber protein lacking one or more amino acids, as herein described, can optionally comprise a non-native residue (e.g., several non-native amino acids) in addition to (i.e., insertions) or in place of (i.e., substitutions) the  
25 missing native amino acid(s); of course, alternatively, the native amino acid(s) can be deleted from the knob. Preferably, the amino-acid is substituted with another non-native amino acid to preserve topology and, especially, trimerization. Moreover, if substituted, the replacement amino acid preferably confers novel qualities to the recombinant fiber protein. For example, to maximally ablate  
30 binding to the native substrate, a native amino acid can be substituted with a residue (or a plurality of residues) having a different charge. Such a substitution maximally interferes with the electrostatic interaction between native adenoviral knob domains and cellular ARs or interferes with a conformational change required to efficiently bind an AR or elements of the RES. Similarly, a native  
35 amino acid can be substituted with a residue (or a plurality of residues) of differing weight, where possible. For example, substitution with a heavier residue

maximally interferes with the steric interaction between adenoviral domains and native substrates, by virtue of the longer side-chains on such heavier residues.

Any native amino-acid residue mediating or assisting in the interaction between the knob and a native cellular AR is a suitable amino acid for mutation or deletion from the recombinant fiber protein. Such amino acid need not itself be the site of contact between the fiber and the receptor. For example, the native amino acid might be involved in a conformational change associated with receptor binding. The inventive fiber protein can lack any number of such native amino acids, so long as, in the aggregate, the recombinant fiber protein can associate to form a trimer. The amino acid can be within a  $\beta$ -sheet of the knob or within a loop connecting two  $\beta$ -sheets (such as, for example, the AB, BC, CD, DE, EF, FG, GH, HI, or IJ loops). Indeed, the amino acid can be within 10 (e.g., within 5) residues of a  $\beta$  sheet or a loop. In the mature, folded trimer of the present invention, the amino acid can be within about 10 nm (e.g., within about 5 nm or even within about 2 nm) of a  $\beta$  sheet or a loop.

Native amino acid residues for modification or deletion can be selected by any method. For example, the sequences from different adenoviral serotypes (which are known in the art) can be compared to deduce conserved residues likely to mediate AR-binding. Alternatively or in combination, the sequence can be mapped onto a three dimensional representation of the protein (such as the crystal structure) to deduce those residues most likely responsible for AR binding. These analyses can be aided by resorting to any common algorithm or program for deducing protein structural functional interaction. Alternatively, random mutations can be introduced into a cloned adenoviral fiber expression cassette. One method of introducing random mutations into a protein is via the *Taq* polymerase. For example, a clone encoding the fiber knob (see, e.g., Roelvink *et al.*, *J. Virol.*, 70, 7614-21 (1996)) can serve as a template for PCR amplification of the adenoviral fiber knob, or a portion thereof. By varying the concentration of divalent cations in the PCR reaction, the error rate of the transcripts can be largely predetermined (see, e.g., Weiss *et al.*, *J. Virol.*, 71, 4385-94 (1997); Zhou *et al.*, *Nucl. Acid. Res.*, 19, 6052 (1991)). The PCR products then can be subcloned back into the template vector to replace the sequence within the fiber coding sequence employed as a source for the PCR reaction, thus generating a library of fibers, some of which will harbor mutations which diminish native AR binding while retaining the ability to trimerize.

The amino acids of knobs from strains other than Ad5 that correspond to these listed residues are apparent upon a comparison between the sequences of the

fibers of different adenoviral strains, and any suitable method of determining such correspondence can be employed (e.g., Clusal method with PAM100 residue weight table, J. Hain method with PAM 250 residue weight table, etc.). Examples of such sequence comparison of the knobs of Ad fiber proteins (SEQ ID NOs:5-25) are set forth in Figures 1A-2B. By such comparison, residues (e.g., conserved) from other serotypes which, mutated as described, result in fiber trimers with reduced AR binding can be identified (see, e.g., SEQ ID NOs: 29-32). Thus, for example, for CAR-binding fibers, preferably, the amino acid(s) to be mutated is within 10 (e.g., within about 5) amino acids or within about 10 nm (e.g., within about 5 nm) of an amino acid corresponding to residues 404-406, 408, 409, 412-417, 420, 439, 441, 442, 449-454, 456, 458, 460, 462, 466, 467, 469-472, 474-477, 482, 485, 487-492, 505-512, 515, 517, 519, 521-528, 533, 535, 537-549, 551, 553, 555, 559-568, 580, or 581 of the native Ad5 fiber protein (SEQ ID NO:1). More preferably, the amino acid(s) to be mutated correspond to at least one of these residues, such as amino acid 189, 190, 198, 201, or 262 of the native Ad9 fiber protein (SEQ ID NO:3) or amino acid 395, 396, 404, 407, or 470 of the native Ad41 long fiber protein (SEQ ID NO:2). Even more preferably, the mutant fiber protein comprises at least one replacement mutation of a residue corresponding to residues 408, 409, 412-417, 420, 477, or 487-491 of the native Ad5 fiber protein or at least one deletion mutation of a residue corresponding to residues 474-477 or 489-492 of the native Ad5 fiber protein. Similarly, for group B fibers, the amino acid(s) to be mutated is within 10 (e.g., within about 5) amino acids or within about 10 nm (e.g., within about 5 nm) of an amino acid corresponding to residues 136, 155, 177, 181, 198, 210, 211, 215, 233, 234, 236, 238, 248, 257, 260, 261, 276, 284, 302, 303, 317, or 318 of the native Ad3 fiber protein (SEQ ID NO:4).

The recombinant fiber protein of the present invention can be produced by any suitable method. For example, the mutant fiber protein can be synthesized using standard direct peptide synthesizing techniques (e.g., as summarized in Bodanszky, *Principles of Peptide Synthesis* (Springer-Verlag, Heidelberg: 1984)), such as via solid-phase synthesis (see, e.g., Merrifield, *J. Am. Chem. Soc.*, 85, 2149-54 (1963); and Barany *et al.*, *Int. J. Peptide Protein Res.*, 30, 705-739 (1987)). Alternatively, site-specific mutations can be introduced into the recombinant fiber protein by ligating into an expression vector a synthesized oligonucleotide comprising the modified site. Alternatively, a plasmid, oligonucleotide, or other vector encoding the desired mutation can be recombined with the adenoviral genome or with an expression vector encoding the

recombinant fiber protein to introduce the desired mutation. Oligonucleotide-directed site-specific mutagenesis procedures also are appropriate (e.g., Walder *et al.*, *Gene*, 42, 133 (1986); Bauer *et al.*, *Gene*, 37, 73 (1985); Craik, *Biotechniques*, 12-19 (1995); U.S. Patents 4,518,584 (Mark *et al.*) and 4,737,462 (Mark *et al.*)).

5 However engineered, the DNA fragment encoding the recombinant fiber protein can be subcloned into an appropriate vector using well known molecular genetic techniques. The fragment is then transcribed and the peptide subsequently translated *in vitro* within a host cell. Any appropriate expression vector (e.g., Pouwels *et al.*, *Cloning Vectors: A Laboratory Manual* (Elsevier, NY: 1985)) and  
10 corresponding suitable host cells can be employed for production of recombinant peptides. Expression hosts include, but are not limited to, bacterial species, yeast, mammalian or insect host cell systems including baculovirus systems (e.g., Luckow *et al.*, *Bio/Technology*, 6, 47 (1988)), and established cell lines such HEK-293, COS-7, C127, 3T3, CHO, HeLa, BHK, etc. An especially preferred  
15 expression system for preparing modified fibers of the invention is a baculovirus expression system (Wickham *et al.*, *J. Virol.*, 70, 6831-38 (1995)) as it allows the production of high levels of recombinant proteins. Of course, the choice of expression host has ramifications for the type of peptide produced, primarily due to post-translational modification.

20 Once produced, the recombinant fiber proteins are assayed for fiber protein activity. Specifically, the ability of recombinant fiber protein to form trimers, interact with the penton base, and interact with native substrate's (e.g., antibodies, ARs, opsonins, collectins, RES cells, etc.) is assayed. Any suitable assay can be employed to measure these parameters. For example, as improperly folded  
25 monomers are generally insoluble (Scopes, "Protein Purification" (3d Ed., 1994), Chapter 9, p. 270-82 (Springer-Verlag, New York)), one assay for trimerization is whether the recombinant fiber is soluble. Determining solubility of the fiber is aided if an amount of radioactive amino-acid is incorporated into the protein during synthesis. Lysate from the host cell expressing the recombinant fiber  
30 protein can be centrifuged, and the supernatant and pellet can be assayed via a scintillation counter or by Western analysis. Subsequently, the proteins within the pellet and the supernatant are separated (e.g., on an SDS-PAGE gel) to isolate the fiber protein for further assay. Comparison of the amount of fiber protein isolated from the pellet vis-à-vis the fiber protein isolated from the supernatant indicates  
35 whether the mutant protein is soluble. Alternatively, trimerization can be assayed by using a monoclonal antibody recognizing only the amino portion of the trimeric form of the fiber (e.g., via immunoprecipitation, Western blotting, etc.). Another

measure of trimerization is the ability of the recombinant fiber to form a complex with the penton base (Novelli and Boulanger, *Virology*, 185, 1189 (1995)), as only fiber trimers can so interact. This propensity can be assayed by co-immunoprecipitation, gel mobility-shift assays, SDS-PAGE (boiled samples migrate as monomers, otherwise, they migrate as larger proteins), etc. Yet another measure of trimerization is to detect the difference in molecular weight of a trimer as opposed to a monomer. For example, a boiled and denatured trimer will run as a lower molecular weight than a non-denatured stable trimer (Hong and Angler, *J. Virol.*, 70, 7071-78 (1996)). A trimeric recombinant fiber also can be assayed for its ability to bind native substrates. For example, modification of fiber to interfere with its interaction with the host innate or acquired immune system can be accomplished by measuring the free titer of the virus over time. This can be assessed by measuring serum half life, tropism to organs associated with the RES (e.g., liver in mice and humans, lung in pigs, etc.), by agglutination of red blood cells, or by detection of adenoviral genetic material in cell samples.

A trimeric recombinant fiber also can be assayed for its ability to bind native ARs. Any suitable assay that can detect this characteristic is sufficient for use in the present invention. A preferred assay involves exposing cells expressing a native AR (e.g., HEK-293 cells) to the recombinant fiber trimers under standard conditions of infection. Subsequently, the cells are exposed to native adenoviruses, and the ability of the viruses to bind the cells is monitored. Monitoring can be by autoradiography (e.g., employing radioactive viruses), immunocytochemistry, or by measuring the level of infection or gene delivery (e.g., using a reporter gene). In contrast with native trimers which reduce or substantially eliminate subsequent viral binding to the HEK-293 cells, those trimers not substantially reducing the ability of native adenoviruses to subsequently bind the cells are trimers of the present invention. The reduction of interference with subsequent viral binding indicates that the trimer is itself not a ligand for its native mammalian AR, or at least binds with reduced affinity.

Alternatively, a vector including a sequence encoding a mutated fiber (or a library of putative mutated fibers, such as described herein) can be introduced into a suitable host cell strain to express the fiber protein, and, mutants can be identified by assaying the inability to bind the soluble CAR protein (e.g., by probing a replica lift with radiolabeled CAR or by other suitable method). Because a reduction in CAR-binding could be due to either selective ablation of the ligand or structural modification affecting trimerization, mutant fibers

identified as non-CAR binding by such a library screen must be assayed for the ability to trimerize, as described above.

### Virion and Virus

5           The present invention provides an adenoviral virion incorporating the recombinant fiber protein of the present invention. The virion does not interact with native substrates (e.g., innate and acquired immune systems, cell-surface proteins, etc.) as readily as the wild-type serotype, due to the above-mentioned reduction in affinity of the fibers present in the virion. Moreover, the virion can be  
10 further modified to reduce interaction with native substrates through the inclusion of other recombinant proteins. Thus, for example, the virion can include one or more recombinant penton base proteins lacking a native RGD sequence to reduce cell binding via  $\alpha_v$  integrins (see, e.g., U.S. patents 5,559,099 (Wickham *et al.*) and 5,731,190 (Wickham *et al.*)). Similarly, the virion can include one or more  
15 recombinant hexons lacking a native sequence (e.g., HVR) to reduce its ability to be recognized by a neutralizing antibody (see, e.g., International Patent Application WO 98/40509 (Crystal *et al.*)). Also, the virion can be modified to reduce its ability to interact with the RES. For example, the virion proteins can be mutated to lack one or more glycosylation or phosphorylation sites, or it can be  
20 produced in the presence of inhibitors of glycosylation or phosphorylation. Similarly, the virion proteins can be conjugated to a lipid derivative of PEG comprising a primary amine group, an epoxy group, or a diacylglycerol group, as discussed above, to reduce collectin and/or opsonin affinity or scavenging by Kupffer cells or other cells of the RES. Such modifications reduce the ability of  
25 host animals to develop neutralizing antibodies to the virions, thereby permitting repeat administration of the virions.

While the virion exhibits reduced affinity for natural adenoviral substrates, it can include one or more non-adenoviral ligands, for example, to effect targeted infection of a population of cells other than that for which adenoviruses are  
30 naturally tropic. Additionally, the non-native ligand can be used to purify the virus, to inactivate the virus (e.g., by adsorbing it to a substrate for the ligand), or to grow the virus on cell lines having receptors recognizing the non-native ligand, for example, as described in International Patent Application WO 98/54346 (Wickham *et al.*).

35           The virus can include any suitable ligand (e.g., a peptide specifically binding to a substrate). For example, for targeting the adenovirus to a cell type other than that naturally infected (or a group of cell types other than the natural

range or set of host cells), the ligand can bind a cell surface binding site (e.g., any site present on the surface of a cell with which the adenovirus can interact to bind the cell and thereby promote cell entry). A cell surface binding site can be any suitable type of molecule, but typically is a protein (including a modified protein such as a glycoprotein, a mucoprotein, etc.), a carbohydrate, a proteoglycan, a lipid, a mucin molecule, or other similar molecule. Examples of potential cell surface binding sites include, but are not limited to, heparin and chondroitin sulfate moieties found on glycosaminoglycans; sialic acid moieties found on mucins, glycoproteins, and gangliosides; common carbohydrate molecules found in membrane glycoproteins, including mannose, N-acetyl-galactosamine, N-acetyl-glucosamine, fucose, and galactose; glycoproteins such cell adhesion molecules (CAMs) (e.g., ICAM-1, ICAM-2, ICAM-3, VCAM-1, NCAM), selectins (e.g., E-selectin, P-selectin, L-selectin, etc.), CD, cadherins, TNF family receptors, GPI-linked receptors, receptors that are efficiently internalized (e.g., CD44, CD31 on endothelial cells, CD34 on high endo-venules), endoglin, growth factor receptors, PSA, androgen receptors, glucocorticoid receptors, prostate-specific membrane antigen (PSMA), MUC1, MUC234, MUC5AC, MUC5B, MUC7, KSA carcino-embryonic antigen (CEA), HER2/NEU (erbB2), folate receptor, corionic gonadotropin- $\beta$ , (Zhang *et al.*, *Clin. Cancer Res.*, 4, 2669-76 (1998); *Cancer Res.*, 58, 4055 (1998)), and others are known in the art.

A particular cell surface binding site can be present on a narrow class of cell types (e.g., cardiac muscle, skeletal muscle, smooth muscle, etc.) or a broader group encompassing several cell types. Through integration of an appropriate cell-specific ligand, the virion can be employed to target any desired cell type, such as, for example, neuronal, glial, endothelial (e.g., via tissue factor receptor, FLT-1, CD31; CD36; CD34, CD105, CD13, ICAM-1 (McCormick *et al.*, *J. Biol. Chem.*, 273, 26323-29 (1998)); thrombomodulin receptor (Lupus *et al.*, *Suppl.*, 2, S120 (1998)); VEGFR-3 (Lymboussaki *et al.*, *Am. J. Pathol.*, 153(2), 395-403 (1998); mannose receptor; VCAM-1 (Schwarzacher *et al.*, *Atherosclerosis*, 122, 59-67 (1996)), or other receptors); blood clots (e.g., through fibrinogen or aIIb $\beta$ 3 peptide), epithelial (e.g., inflamed tissue through selectins, VCAM-1, ICAM-1, etc.), keratinocytes, follicular cells, adipocytes, fibroblasts, hematopoietic or other stem cells, myoblasts, myofibers, cardiomyocytes, smooth muscle, somatic, osteoclasts, osteoblasts, tooth blasts, chondrocytes, melanocytes, hematopoietic cells, etc., as well as cancer cells derived from any of the above cell types (e.g., prostate (such as via PSMA receptor (see, e.g., Schuur *et al.*, *J. Biol. Chem.*, 271, 7043 (1998); *Cancer Res.*, 58, 4055 (1998))), breast, lung, brain (e.g.,

glioblastoma), leukemia/lymphoma, liver, sarcoma, bone, colon, testicular, ovarian, bladder, throat, stomach, pancreas, rectum, skin (e.g., melanoma), kidney, etc.). Thus, the inventive virions can be targeted to cells within any organ or system, including, for example, respiratory system (e.g., trachea, upper airways, lower airways, alveoli), nervous system and sensory organs (e.g., skin, ear, nasal, tongue, eye), digestive system (e.g., oral epithelium and sensory organs, salivary glands, stomach, small intestines/duodenum, colon, gall bladder, pancreas, rectum), muscular system (e.g., skeletal muscle, connective tissue, tendons), skeletal system (e.g., joints (synovial cells), osteoclasts, osteoblasts, etc.), immune system (e.g., bone marrow, stem cells, spleen, thymus, lymphatic system, etc.), circulatory system (e.g., muscles connective tissue, and/or endothelia of the arteries, veins, capillaries, etc.), reproductive system (e.g., testis, prostate, uterus, ovaries), urinary system (e.g., bladder, kidney, urethra), endocrine or exocrine glands (e.g., breasts, adrenal glands, pituitary glands), etc.

In other embodiments (e.g., to facilitate purification or propagation within a specific engineered cell type), the non-native ligand can bind a compound other than a cell-surface protein. Thus, the ligand can bind blood- and/or lymph-borne proteins (e.g., albumin), synthetic peptide sequences such as polyamino acids (e.g., polylysine, polyhistidine, etc.), artificial peptide sequences (e.g., FLAG), and RGD peptide fragments (Pasqualini *et al.*, *J. Cell. Biol.*, 130, 1189 (1995)). The ligand can even bind non-peptide substrates, such as plastic (e.g., Adey *et al.*, *Gene*, 156, 27 (1995)), biotin (Saggio *et al.*, *Biochem. J.*, 293, 613 (1993)), a DNA sequence (Cheng *et al.*, *Gene*, 171, 1 (1996); Krook *et al.*, *Biochem. Biophys. Res. Commun.*, 204, 849 (1994)), streptavidin (Geibel *et al.*, *Biochemistry*, 34, 15430 (1995); Katz, *Biochemistry*, 34, 15421 (1995)), nitrostreptavidin (Balass *et al.*, *Anal. Biochem.*, 243, 264 (1996)), heparin (Wickham *et al.*, *Nature Biotechnol.*, 14, 1570-73 (1996)), cationic supports, metals such as nickel and zinc (e.g., Rebar *et al.*, *Science*, 263, 671 (1994); Qui *et al.*, *Biochemistry*, 33, 8319 (1994)), or other potential substrates.

Examples of suitable ligands and their substrates for use in the method of the invention include, but are not limited to, CR2 receptor binding the amino acid residue attachment sequences, CD4 receptor recognizing the V3 loop of HIV gp120, transferrin receptor and its ligand (transferrin), low density lipoprotein receptor and its ligand, the ICAM-1 receptor on epithelial and endothelial cells in lung and its ligand, linear or cyclic peptide ligands for streptavidin or nitrostreptavidin (Katz, *Biochemistry*, 34, 15421 (1995)), galactin sequences that bind lactose, galactose and other galactose-containing compounds, and



asialoglycoproteins that recognize deglycosylated protein ligands. Moreover, additional ligands and their binding sites preferably include (but are not limited to) short (e.g., 6 amino acids or less) linear stretches of amino acids recognized by integrins, as well as polyamino acid sequences such as polylysine, polyarginine, etc. Inserting multiple lysines and/or arginines provides for recognition of heparin and DNA. Also, a ligand can comprise a commonly employed peptide tag (e.g., short amino acid sequences known to be recognized by available antisera) such as sequences from glutathione-S-transferase (GST) from *Shistosoma manosi*, thioredoxin  $\beta$ -galactosidase, or maltose binding protein (MPB) from *E. coli*, human alkaline phosphatase, the FLAG octapeptide, hemagglutinin (HA) (Wickham *et al.* (1996), *supra*), polyoma virus peptides, the SV40 large T antigen peptide, BPV peptides, the hepatitis C virus core and envelope E2 peptides and single chain antibodies recognizing them (Chan, *J. Gen. Virol.*, 77, 2531 (1996)), the c-myc peptide, adenoviral penton base epitopes (Stuart *et al.*, *EMBO J.*, 16, 1189-98 (1997)), epitopes present in the E2 envelope of the hepatitis C virus (see, e.g., Chan *et al.* (1996), *supra*), and other commonly employed tags. A preferred substrate for a tag ligand is an antibody directed against it or a derivative of such an antibody (e.g., a FAB fragment, single chain antibody (ScAb)).

As mentioned, a suitable ligand can be specific for any desired substrate, such as those recited herein or otherwise known in the art. However, adenoviral vectors also can be engineered to include novel ligands (e.g., in protein II, III, IIIa, IV, IV, VI, and/or IX) by first assaying for the ability of a peptide to interact with a given substrate. Generally, a random or semirandom peptide library containing potential ligands can be produced, which is essentially a library within an expression vector system. Such a library can be screened by exposing the expressed proteins (i.e., the putative ligands) to a desired substrate. Positive selective binding of a species within the library to the substrate indicates a ligand for that substrate, at least under the conditions of the assay. For screening such a peptide library, any assay able to detect interactions between proteins and substrates is appropriate, and many are known in the art. However, one preferred assay for screening a protein library is a display system (e.g., using an adenovirus or a bacteriophage), which employs a virus expressing the library (e.g., Koivunen *et al.*, *Bio/Technology*, 13, 265-70 (1995); Yanofsky *et al.*, *Proc. Nat. Acad. Sci. U.S.A.*, 93, 7381-86 (1996); Barry *et al.*, *Nature Med.*, 2(3), 299-305 (1996)). Binding of the virus to the substrate is assayed by exposing the virus to the substrate, rinsing the substrate, and selecting for virus remaining bound to the substrate. Subsequently, limiting dilution can identify individual clones

expressing the putative ligand. Thereafter, the insert present in such clones can be sequenced to determine the identity of the ligand.

Once a given ligand is identified, it can be incorporated into any location of the virus capable of interacting with a substrate (i.e., the viral surface). For example, the ligand can be incorporated into the fiber, the penton base, the hexon, protein IX, VI, or IIIa, or other suitable location. Where the ligand is attached to the fiber protein, preferably it does not disturb the interaction between viral proteins or monomers. Thus, the ligand preferably is not itself an oligomerization domain, as such can adversely interact with the trimerization domain as discussed above. Preferably the ligand is added to the virion protein, and is incorporated in such a manner as to be readily exposed to the substrate (e.g., at the terminus of the protein, attached to a residue facing the substrate, positioned on a peptide spacer to contact the substrate, etc.) to maximally present the ligand to the substrate. Where the ligand is attached to or replaces a portion of the penton base, preferably it is within the hypervariable regions to ensure that it contacts the substrate. Furthermore, where the ligand is attached to the penton base, preferably, the recombinant fiber is truncated or short (e.g., from 0 to about 10 shaft repeats) to maximally present the ligand to the substrate (see, e.g., U.S. Patent 5,559,099 (Wickham *et al.*)). Where the ligand is attached to the hexon, preferably it is within a hypervariable region (Mikszta *et al.*, *J. Virol.*, 70(3), 1836-44 (1996)).

When engineered into an adenoviral protein, the ligand can comprise a portion of the native sequence in part and a portion of the non-native sequence in part. Similarly, the sequences (either native and/or nonnative) that comprise the ligand in the protein need not necessarily be contiguous in the chain of amino acids that comprise the protein. In other words, the ligand can be generated by the particular conformation of the protein, e.g., through folding of the protein in such a way as to bring contiguous and/or noncontiguous sequences into mutual proximity. Of course an adenovirus of the present invention (or a blocking protein) can comprise multiple ligands, each binding to a different substrate. For example, a virus can comprise a first ligand permitting affinity purification as described herein, a second ligand that selectively binds a cell-surface site as described herein, and/or a third ligand for inactivating the virus, also as described herein.

The protein including the ligand can include other non-native elements as well. For example, a non-native, unique protease site also can be inserted into the amino acid sequence. The protease site preferably does not affect fiber trimerization or substrate specificity of the fiber ligand. Many such protease sites are known in the art. For example, thrombin recognizes and cleaves at a known

amino acid sequence (Stenflo *et al.*, *J. Biol. Chem.*, 257, 12280-90 (1982)). The presence of such a protease recognition sequence facilitates purification of the virus in some protocols. The protein can be engineered to include the ligand by any suitable method, such as those methods described above for introducing mutations into proteins.

The virion can be used by itself, for example in studies of viral tropism or binding kinetics. In other embodiments, the virion can be used as a genetic vector. For example, the virion can be used in conjunction with lipids and/or liposomes to deliver exogenous genetic material to target cells, in accordance with well-documented methods. In other embodiments, the virion contains a genome derived from an adenovirus; thus, the invention provides an adenoviral vector including the inventive virion and an adenoviral genome.

The adenoviral vector of the present invention can include one or more non-native amino acid sequences for expression (e.g., "expression cassettes" or "genes") as well. Preferably, the non-native amino acid is capable of being transcribed in a cell into which the vector has been internalized. The non-native amino acid can encode a product that effects a biological (e.g., therapeutic) response either at the cellular level or systemically; alternatively, the non-native nucleic acid sequence can encode a product that, in some fashion, can be detected in a cell (e.g., a "reporter gene"). The non-native amino acid can exert its effect at the level of RNA or protein. For instance, a protein encoded by the non-native amino acid can be employed in the treatment of an inherited disease, such as, e.g., the cystic fibrosis transmembrane conductance regulator cDNA for the treatment of cystic fibrosis. Alternatively, the protein encoded by the non-native amino acid can exert its therapeutic effect by effecting cell death. For instance, expression of the non-native amino acid in itself can lead to cell killing, as with expression of the diphtheria toxin. Alternatively, the expression of the non-native amino acid, can render cells selectively sensitive to the action of certain drugs, e.g., expression of the HSV thymidine kinase gene renders cells sensitive to antiviral compounds including acyclovir, gancyclovir, and FIAU (1-(2-deoxy-2-fluoro- $\beta$ -D-arabinofuranosil)-5-iodouracil). Moreover, the non-native amino acid can exert its effect at the level of RNA, for instance, by encoding an antisense message or ribozyme, a protein which affects splicing or 3' processing (e.g., polyadenylation), or a protein affecting the level of expression of another gene within the cell (i.e., where gene expression is broadly considered to include all steps from initiation of transcription through production of a processed protein), perhaps, among other things, by mediating an altered rate of mRNA accumulation, an alteration of

mRNA transport, and/or a change in post-transcriptional regulation. Of course, where it is desired to employ gene transfer technology to deliver a given non-native amino acid, its sequence will be known in the art.

Where the inventive adenoviral vector includes a non-native amino acid  
5 and a non-adenoviral ligand in its virion, the non-native amino acid can be operably linked to any suitable promoter, such as a promoter native to the adenoviral genome or a non-adenoviral promoter. Where the ligand is employed to deliver the vector to a desired cell type, preferably the non-adenoviral promoter is active within the cell type, and more preferably, the non-adenoviral promoter is  
10 a tissue-specific promoter (e.g., specific for the cell type to which the ligand binds), such as those cell types discussed above. For example, expression in targeted endothelial cells can be mediated using the E-selectin promoter (see, e.g., Whelan *et al.*, *TIBS*, 21, 65-69 (1996)); passenger gene expression in targeted prostate cancer cells can be mediated using the PSA promoter (see, e.g., Schuur *et al.*, *J. Cell Biol.*, 271, 7043 (1996). Pang *et al.*, *Cancer Res.*, 57, 495 (1997)) or the  
15 E2F promoter. Furthermore, the promoter can be that for a tissue-specific receptor, such as those receptors mentioned herein, still other tissue specific promoter systems are known in the art. Alternatively, the non-native amino acid can be placed under control of a regulable promoter (e.g., metallothionein  
20 promoter, tetracycline-responsive promoter, RU486-responsive promoter, etc.).

The altered protein (e.g., the recombinant fiber protein or the coat protein having the ligand) and the non-native amino acid where present) can be incorporated into the adenovirus by any suitable method, many of which are known in the art. As mentioned herein, the protein is preferably identified by  
25 assaying products produced in high volume from genes within expression vectors (e.g., baculovirus vectors). The genes from the vectors harboring the desired mutation can be readily subcloned into plasmids, which are then transfected into suitable packaging cells (e.g., HEK-293 cells). Transfected cells are then incubated with adenoviruses under conditions suitable for infection. At some  
30 frequency within the cells, homologous recombination between the vector and the virus will produce an adenoviral genome harboring the desired mutation.

Adenoviruses of the present invention can be either replication competent or replication deficient. Preferably, the adenoviral vector comprises a genome with at least one modification therein, rendering the virus replication deficient  
35 (see, e.g., International Patent Application WO 95/34671 (Kovesdi *et al.*)). The modification to the adenoviral genome includes, but is not limited to, addition of a DNA segment, rearrangement of a DNA segment, deletion of a DNA segment.

replacement of a DNA segment, or introduction of a DNA lesion. A DNA segment can be as small as one nucleotide and as large as the adenoviral genome (e.g., about 36 kb) or, alternately, can equal the maximum amount which can be packaged into an adenoviral virion (i.e., about 38 kb). Preferred modifications to the adenoviral genome include modifications in the E1, E2, E3, and/or E4 regions. An adenovirus also preferably can be a cointegrate, i.e., a ligation of adenoviral genomic sequences with other sequences, such as other virus, phage, or plasmid sequences.

The virion and adenoviral vector of the present invention have many qualities which render them attractive choices for use in gene transfer, as well as other applications. For example, in many embodiments, the adenovirus does not infect its native host cells as readily as does wild-type adenovirus, due to the recombinant fiber protein. Moreover, by virtue of additional modifications, such virions and vectors are less readily cleared from the host by the innate or acquired immune responses, thus boosting effective free titer and lengthening serum half-life. Furthermore, the virions and vectors have at least one non-native ligand specific for a substrate which facilitates viral propagation, targeting, purification, and/or inactivation as discussed herein. The presence of such ligands can effectively confine expression of non-native amino acids within a predefined cell type or tissue. Linking the non-native amino acid to a tissue-specific or regulable promoter further minimizes expression of the non-native amino acid outside of the targeted tissue. The ligands and the trimerization domains can be separate domains, thus permitting the virus to be easily be reengineered to incorporate different ligands without perturbing fiber trimerization.

Of course, for delivery into a host (such as an animal), a virus of the present invention can be incorporated into a suitable carrier. As such, the present invention provides a composition comprising an adenovirus of the present invention and a pharmacologically acceptable carrier (e.g., a pharmaceutically-acceptable carrier). Any suitable preparation is within the scope of the invention. The exact formulation, of course, depends on the nature of the desired application (e.g., cell type, mode of administration, etc.), and many suitable preparations are set forth in U.S. Patent 5,559,099 (Wickham *et al.*).

### Cell Line

As mentioned herein, an adenovirus of the present invention does not readily infect its native host cell via the native AR because its ability to bind ARs is significantly attenuated (due to the incorporation of the recombinant fiber

protein of the present invention). Therefore, the invention provides a cell line able to propagate the inventive adenovirus. Preferably, the cell line can support viral growth for at least about 10 passages (e.g., about 15 passages), and more preferably for at least about 20 passages (e.g., about 25 passages), or even 30 or more passages.

For example, the adenoviruses can be first grown in a packaging cell line which expresses a native fiber protein gene. The resultant viral particles are therefore likely to contain both native fibers encoded by the complementing cell line and non-native fibers encoded by the adenoviral genome (such as those fibers described herein): hence a population of such resultant viruses will contain both fiber types. Such particles will be able to bind and enter packaging cell lines via the native fiber more efficiently than particles which lack native fiber molecules. Thus, the employment of such a fiber-encoding cell line permits adenovirus genomes encoding chimeric, targeted adenovirus fibers to be grown and amplified to suitably high titers. The resultant "mixed" stocks of adenovirus produced from the cell lines encoding the native fiber molecule will contain both native and chimeric adenovirus fiber molecules; however, the particles contain genomes encoding only the chimeric adenovirus fiber. Thus, to produce a pure stock of adenoviruses having only the chimeric adenovirus fiber molecules, the "mixed" stock is used to infect a packaging cell line which does not produce native fiber (such as HEK-293 for E1-deleted non-group B viruses). The resultant adenoviruses contain only the fiber molecules encoded by the genomes (i.e., the chimeric fiber molecules).

Similar fiber-complementing cell lines have been produced and used to grow mutant adenovirus lacking the fiber gene). However, the production rates of these cell lines have generally not been great enough to produce adenovirus titers of the fiber-deleted adenovirus comparable to those of fiber-expressing adenovirus particles. The lower titers produced by such mutants can be improved by temporally regulating the expression of the native fiber to more fully complement the mutant adenovirus genome. One strategy to produce such an improved cell line is to use of a regulable promoter to permit fiber production to be controlled and activated once the cells are infected with adenovirus. Alternatively, an efficient mRNA splice site introduced into the fiber gene in the complementing cell line improves the level of fiber protein production in the cell line.

When the adenovirus is engineered to contain a ligand specific for a given cell surface binding site, any cell line expressing that receptor and capable of supporting adenoviral growth is a suitable host cell line. However, because many

ligands do not bind cell surface binding sites (especially some of the novel ligands discussed herein), a cell line can be engineered to express the substrate for the ligand.

The present invention provides a cell line expressing a non-native cell-  
5 surface receptor (a pseudo-receptor) to which a virus having a ligand for said receptor binds. Any cell line capable of supporting viral growth is a suitable cell line for use in the present invention. If the virus lacks genes essential for viral replication, preferably the cell line expresses complementing levels of such gene products (see, e.g., International Patent Application WO 95/34671 (Kovesdi et al.),  
10 U.S. Patents 5,658,724 (DeLuca) and 5,804,413 (DeLuca)). When the virus is an adenovirus, preferably the cell line of the present invention is derived from HEK-293 cells. When the virus is a herpesvirus, preferably the cell line of the present invention is derived from VERO cells.

The non-native cell surface binding site is a substrate molecule, such as  
15 described herein, to which an adenovirus having a ligand selectively binding that substrate can bind the cell and thereby promote cell entry. The binding site can recognize a non-native ligand incorporated into the adenoviral coat or a ligand native to a virus. For example, when the non-native viral ligand is a tag peptide, the binding site can be a single chain antibody (ScAb) receptor recognizing the  
20 tag. Alternatively, the ScAb can recognize an epitope present in a region of a mutated fiber knob (if present), or even an epitope present on a native adenoviral coat protein, (e.g., on the fiber, penton, hexon, etc.). Alternatively, if the non-native ligand recognizes a cell-surface substrate (e.g., membrane-bound protein), the binding site can comprise that substrate. If the substrate binding site is native  
25 to a cell-surface receptor, the cell line can express a mutant receptor with decreased ability to interact with the cellular signal transduction pathway (e.g., a truncated receptor, such as NMDA (Li et al., *Nat. Biotech.*, 14, 989 (1996))), attenuated ability to act as an ion channel, or other modification. Infection via such modified proteins minimizes the secondary effects of viral infection on host-  
30 cell metabolism by reducing the activation of intracellular messaging pathways and their various response elements. The choice of binding site depends to a large extent on the nature of the adenovirus. However, to promote specificity of the virus for a particular cell type, the binding site preferably is not a native mammalian AR. Moreover, the binding site must be expressed on the surface of  
35 the cell to be accessible to the virus. Hence, where the binding site is a protein, it preferably has a leader sequence and a membrane tethering sequence to promote

proper integration into the membrane (see, e.g., Davitz, *et al.*, *J. Exp. Med.* 163, 1150 (1986)).

The cell line can be produced by any suitable method. For example, a vector (e.g., an oligonucleotide, plasmid, viral, or other vector) containing a nucleic acid encoding the non-native receptor can be introduced into source cell line by conventional means. Preferably, the vector also encodes an agent permitting the cells harboring it to be selected (e.g., the vector can encode resistance to antibiotics which kill cells not harboring the plasmid). At some frequency, the vector will recombine with the cell genome to produce a transformed cell line expressing the non-native receptor.

### EXAMPLES

While it is believed that one of skill in the art is fully able to practice the invention after reading the foregoing description, the following examples further illustrate some of its features. In particular, the examples demonstrate the construction of several recombinant fiber proteins, each exhibiting reduced affinity for native adenoviral substrates. The examples further demonstrate the inclusion of such recombinant fiber proteins into adenoviral vectors, and the retargeting of such vectors using non-native ligands. The examples also demonstrate the successful construction of a pseudoreceptor cell line able to propagate the alternatively targeted viruses. As these examples are included for purely illustrative purposes, they should not be construed to limit the scope of the invention in any respect.

The procedures employed in these examples, such as affinity chromatography, Southern blots, PCR, DNA sequencing, vector construction (including DNA extraction, isolation, restriction digestion, ligation, etc.), cell culture (including antibiotic selection), transfection of cells, protein assays (Western blotting, immunoprecipitation, immunofluorescence), etc., are techniques routinely performed by those of skill in the art (see generally Sambrook *et al.*, *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989)). Accordingly, in the interest of brevity, experimental protocols are not discussed in detail.



**EXAMPLE 1**

This example describes mutant fiber trimers exhibiting reduced affinity for the CAR protein.

Using standard site-directed mutagenesis, mutations were introduced into nearly every major sheet and loop in the native Ad5 fiber knob sequence (SEQ ID NO:1). In a first series of mutagenesis, replacement mutations were designed in which between 3 and 6 contiguous amino acids within a loop were replaced by the same number of glycine residues. In a second series of mutagenesis, mutations were designed in which between 1 and 4 amino acids were deleted from the native sequence. Extensive point mutations also were conducted. One additional mutant was designed in which 12 amino acids were deleted and replaced with a tetrapeptide sequence.

Respective baculovirus clones, each containing one of the recombinant mutant protein genes, were created and used to produce recombinant mutant knob proteins in insect cells. The baculovirus-infected insect cells were freeze-thawed at 3 days post-infection to release any soluble recombinant mutant protein (approximately  $10^7$  cells per ml of PBS). The freeze-thawed lysate was centrifuged and the soluble fraction and the insoluble pellet were collected. Western analysis of the soluble and insoluble fractions revealed that similar levels of the mutant and native fiber knobs were present in the soluble fraction. Mutants which retained solubility represent proteins which folded properly and trimerized, and these are set forth in Table 1. In the table, mutations are indicated by noting the location of the mutated residue or residues of the Ad5 fiber within parentheses. The identity of the native residue or residues is set forth to the left, and the identity of any substituting residue or residues is to the right of the parentheses. Deletions are further delineated using the " $\Delta$ " symbol.

Table 1

Mutation Location	Mutations	
AB Loop (403-418)	T(404)G P(405)G A(406)K S(408)E S(408)G P(409)A R(412)G	RLN(412-414)GGG N(414)G A(415)G RLNAEK(415-417)SLNGGG E(416)G K(417)G K(417)L
B Sheet (419-428)	K(420)A	
C Sheet (431-440)	L(439)S	
CD Loop (441-453)	V(441)S $\Delta$ SG(449-450)	SGTVQ(449-453)GSGSG
D Sheet (454-461)	S(454)N + R(460)Q H(456)E + R(460)E	I(458)E + R(460)E
DE Loop (462-478)	D(462)A V(466)S L(467)S NNS(469-471)GGG $\Delta$ F(472)	DPE(474-476)GGG DPEY(474-477)GGGG Y(477)A Y(477)T
E Sheet (479-482)	N(482)A	
F Sheet (485-486)	L(485)G	
FG Loop (487-514)	E(487)G T(489)G A(490)G EGTAY(487-491)GGGGG Y(491)A $\Delta$ TAYT(489-492)	P(505)G $\Delta$ K(506) H(506)A $\Delta$ KT(510-511) SHGKTA(507-512)GSGSGS
G Sheet (515-521)	N(515)S + V(517)S V(517)S + Q(519)S	Y(521)H
GH Loop (522-528)	NGDKT(523-527)GSGSG D(525)K KTK(526-528)RSR	K(526)E K(528)S
H Sheet (528-536)	T(535)E	T(533)S + T(535)S
HI Loop (537-549)	N(537)E	GTQETGDTTPSA(538-549)GSGG
I Sheet (550-557)	S(551)N + S(555)N S(551)E	S(553)E
IJ Loop (558-572)	SGHN(559-562)GSGS $\Delta$ HN(561-562) Y(563)H	INEI(564-567)GSGS E(566)K F(568)H
C-Terminus (573-578)	Q(580)G	E(581)G

To determine whether a given mutant fiber had reduced affinity for CAR, competition experiments were performed by preincubating A549 cells with either the trimeric mutants or native fiber knobs followed by incubation with radiolabeled Ad5 virus. Either 1 or 10  $\mu$ l volumes of the native knob preincubated with A549 cells blocked 90% or more of the labeled Ad5 binding to cells measured in the absence of a competitor. In this assay, any soluble, trimeric mutant less efficient in blocking fiber-mediated Ad5 cell binding or gene transduction than the native knob was considered to have reduced affinity for CAR. Those trimeric mutant fibers exhibiting reduced affinity for CAR in this assay are indicated in Table 2.

The trimeric mutant fiber proteins were mass produced by infecting roughly 15 million insect cells each with the baculoviral vectors (MOI = 10) and culturing them for 3 days. The cells were harvested and freeze-thawed, and the cell debris was removed via centrifugation. NaCl was added to the supernatant to a final concentration of 750 mM, and then the supernatant was added to 500  $\mu$ l TALON™ resin. After one hour at 25 °C, the resin was centrifuged at 2,500 for two minutes. The supernatant was removed, and the resin resuspended in 10 ml 750 mM NaCl. After 30 minutes incubation, the resin suspension was run through a column. The mutant protein was eluted using 2 ml of elution fluid (20 mM TRIS, pH 8.0, 100 mM NaCl, 150 mM imidazole). The eluate was dialized once against PBS with 750 mM NaCl, once against PBS with 500 mM NaCl, and once against PBS with 250 mM NaCl. Protein concentration was determined by standard methods and protein integrity verified by Western analysis.

The purified proteins were subjected to a competition assay with Ad5 capsids to assess the degree to which each mutation decreased interaction with CAR. Serial dilutions of each mutant protein, as well as wild-type Ad5 fiber, were added to A549 cells ( $10^5$  cells/well) in 24-well plates. Following this preincubation, an Ad5 vector containing the lacZ gene were added to each well (MOI = 10). After a one hour incubation at 37 °C, the inoculum was removed, the cells were washed with culture medium, and then a culture medium (DMEM with 5 % FCS) added. The cells were incubated overnight, lysed 18 hours post infection, and assayed for  $\beta$ -galactosidase activity by standard methods. Plotting the degree of  $\beta$ -galactosidase activity against concentration of preincubation protein permitted assessment of each protein's  $IC_{50}$  value (the concentration of the competing protein at the 50% level). The degree to which each mutation reduced CAR-binding as calculated by this method is set forth in Table 2.

Table 2

Mutation Number	Mutation Location	Mutation Sequence	Competition
F5K	-	Native	100%
F3K	-	Native	< 0.1%
Ad5-1	AB Loop	S408E	< 0.1%
Ad5-2	AB Loop	P409A	< 1%
Ad5-3	AB Loop	RLNAEK(412-417)SLNGGG	< 0.1%
Ad5-4	AB Loop	K(417)G	< 0.1%
Ad5-5	B Sheet	K(420)A	< 0.1%
Ad5-6	DE Loop	$\Delta$ DPE(474-476)	< 20%
Ad5-7	DE Loop	$\Delta$ DPEY(474-477)	< 0.1%
Ad5-8	DE Loop	Y(477)A	< 0.1%
Ad5-9	FG Loop	EGTAY(487-491)GGGGG	< 0.1%
Ad5-10	FG Loop	$\Delta$ TAYT(489-492)	< 0.1%

**EXAMPLE 2**

This example describes recombinant fiber proteins exhibiting reduced  
5 affinity for the CAR protein.

The Ad9 and long Ad41 fiber proteins corresponding to mutations Ad5-1, Ad5-2, Ad5-4, Ad5-5, and Ad5-9 (see Figures 1A and 1B) were generated. The resultant mutant proteins were soluble, and each was used in competition assays  
10 against wild type Ad5, as described in Example 1, to assess whether the mutations affected CAR binding. The results of these experiments (presented in Table 3) reveal that residues important for CAR binding are conserved among adenoviral serotypes.

Table 3

Mutation	Mutation Sequence	Corresponding Ad5 Mutation	Competition
Ad9-1	S(189)E	Ad5-1	No
Ad9-2	P(190)A	Ad5-2	No
Ad9-3	K(198)G	Ad5-4	No
Ad9-4	K(201)A	Ad5-5	No
Ad9-5	Y(262)A	Ad5-8	No
Ad41-1	S(395)E	Ad5-1	No
Ad41-2	P(369)A	Ad5-2	No
Ad41-3	L(404)G	Ad5-4	No
Ad41-3	T(470)A	Ad5-8	No

**EXAMPLE 3**

This example describes the production of a pseudo-receptor for constructing a cell line able to replicate adenoviruses lacking native cell-binding function (but targeted for the pseudo-receptor). Specifically, the exemplary pseudo-receptor includes a binding domain from a single-chain antibody recognizing HA.

Anti-HA ScFv was constructed as an N-Term-VL-VH fusion protein. RT-PCR was performed on RNA obtained from hybridomas producing HA antibodies using primers specific for  $\kappa$ - or  $\gamma 2\beta$ - and C-terminus of the VL and VH genes (see Gilliland *et al.*, *Tissue Antigens*, 47, 1-20 (1996)). After sequencing the resulting PCR products, specific oligonucleotides were designed to amplify the VL-VH fusion in a second round of PCR. The final PCR product was cloned to create a plasmid for production of anti-HA ScFv in *E. coli*. The expressed protein has a C-terminal E peptide for detection of binding to HA-tagged penton base via Western analysis of ELISA assay. Upon transformation of bacterial cells with the plasmid, Western analysis using an antibody recognizing the E peptide revealed a protein of the expected size.

To determine whether the anti-HA ScFv was functional, it was used in protein A immunoprecipitation assays using adenoviral coat proteins (recombinant penton base) containing the HA epitope. The anti-HA ScFv was able to precipitate

HA-containing penton base proteins. These results indicate the successful construction of the extracellular portion of a pseudo-receptor for binding an adenovirus having a non-native ligand (i.e., HA).

To create an entire anti-HA pseudo-receptor, the anti-HA ScFv was cloned  
5 in frame with sequences encoding a C-terminal pair of myc epitopes followed by the PDGF receptor transmembrane anchor. The entire sequence of this pseudo-receptor is indicated at SEQ ID NO:28. A eukaryotic expression plasmid containing this sequence, pSc(HA), was transfected into HEK-293 cells. The following day the pSc(HA)-transfected cells or cells transfected with a control  
10 ScFv construct were incubated for 30 min on ice with a fluorescein-tagged HA peptide (HA\*) or with a fluorescein-tagged scrambled HA peptide (scrHA\*). Following the incubation of HA\* with the pSc(HA)-transfected cells, a discrete population of cells was found to brightly fluoresce specifically around the cell membrane. The pSc(HA)-transfected cells incubated with the scrHA\* peptide did  
15 not display this fluorescent pattern, nor did the cells transfected with the control plasmid and then incubated with HA\*. Enhanced fluorescence of the pSc(HA)-transfected cells incubated with HA\* was also demonstrated by FACS analysis. Moreover, preincubation of the anti-HA pseudo-receptor cells with excess unlabelled HA peptide, but not unlabelled FLAG peptide, blocked the fluorescent  
20 pattern observed on cells incubated with HA\* alone.

These results demonstrate the successful construction and expression of a cell line consisting essentially of cells expressing a functional pseudo-receptor.

#### EXAMPLE 4

25 This example describes an alternatively targeted adenovirus having recombinant fiber proteins exhibiting reduced affinity for the CAR protein and having a non-native ligand.

The Ad5-10 mutant described in Example 1 was subjected to further site directed mutagenesis to introduce a polypeptide including the HA epitope into the  
30 HI loop of the fiber knob (between amino acids 543 and 544 of SEQ ID NO:1). The resultant fiber has the TAYT deletion in the FG loop and an HA epitope sequence inserted into the HI loop.

The gene encoding this mutant fiber was combined into a plasmid that contains a full length, E1- and E3-deleted adenovirus genome carrying the above  
35 fiber mutation plus a CMV-driven *LacZ* reporter gene in the E1 region. This plasmid was then linearized and transfected into HEK-293 cells expressing the anti-HA pseudo-receptor described in Example 3. After 5 days the cells were

freeze-thawed three times, and the virus-containing lysate was passaged onto fresh anti-HA-293 cells.

The resultant adenoviruses were further amplified in the anti-HA-293 cells and then purified using standard methods. The vector (AdZ.F\*fg(HA)hi) exhibits  
5 reduced binding capacity to CAR on standard HEK-293 cells due to the TAYT deletion; however, it binds with high affinity via its HA epitope to the anti-HA pseudoreceptor present on the anti-HA-293 cell line.

### EXAMPLE 5

10 This example describes an alternatively targeted adenovirus having recombinant fiber proteins exhibiting reduced affinity for the CAR protein and having more than one non-native ligand.

The Ad5-10 mutant described in Example 1 was subjected to further site directed mutagenesis to introduce a polypeptide including the HA epitope and a  
15 high affinity RGD ligand into the HI loop of the fiber knob (between amino acids 543 and 544 of SEQ ID NO:1). The resultant plasmid encodes a fiber with the TAYT deletion in the FG loop and an RGD sequence inserted into the HI loop.

The gene encoding this mutant fiber gene was then combined into a plasmid that contains a full length, E1 and E3-deleted adenovirus genome carrying  
20 the above fiber mutation plus a CMV-driven *LacZ* reporter gene in the E1 region. This plasmid was then linearized and transfected into HEK-293 cells expressing the anti-HA pseudo-receptor described in Example 2. After 5 days the cells are freeze-thawed three times and the virus-containing lysate is passaged onto fresh HEK-293 cells.

25 The resultant adenoviruses were further amplified in the anti-HA-293 cells and then purified using standard methods. The vector exhibits reduced binding capacity to CAR on standard HEK-293 cells due to the TAYT deletion; however, it efficiently infects cells expressing  $\alpha_v$  integrins (such as tumor cells) via the RGD ligand present in the HI loop.

30

### EXAMPLE 6

This example describes an alternatively targeted adenovirus having recombinant fiber proteins exhibiting reduced affinity for the CAR protein and having a non-native ligand.

35 The Ad5-3 mutant described in Example 1 was subjected to further site directed mutagenesis to introduce an 18 amino acid polypeptide including the HA epitope into the HI loop of the fiber knob (between amino acids 543 and 544 of

SEQ ID NO:1) . The resultant fiber has the RLNAEK mutation of the AB loop and an HA epitope sequence inserted into the HI loop.

5 The gene encoding this mutant fiber was combined into a plasmid that contains a full length, E1- and E3-deleted adenovirus genome carrying the above fiber mutation plus a CMV-driven *LacZ* reporter gene in the E1 region. This plasmid was then linearized and transfected into HEK-293 cells expressing the anti-HA pseudo-receptor described in Example 3. After 5 days the cells were freeze-thawed three times, and the virus-containing lysate was passaged onto fresh anti-HA 293 cells.

10 The resultant adenoviruses were further amplified in the anti-HA 293 cells and then purified using standard methods. The vector (AdZ.F\*ab(HA)hi) exhibits reduced binding capacity to CAR on standard HEK-293 cells due to the RLNAEK mutation; however, it binds with high affinity via its HA epitope to the anti-HA pseudoreceptor present on the anti-HA 293 cell line.

15

#### EXAMPLE 7

This example describes an alternatively targeted adenovirus having recombinant fiber proteins exhibiting reduced affinity for the CAR protein and having more than one non-native ligand.

20 The Ad5-3 mutant described in Example 1 was subjected to further site directed mutagenesis to introduce a polypeptide including the HA epitope and a high affinity RGD ligand into the HI loop of the fiber knob (between amino acids 543 and 544 of SEQ ID NO:1) . The resultant plasmid encodes a fiber with the RLNAEK mutation of the AB loop and an HA epitope and RGD sequence inserted into the HI loop.

25 The gene encoding this mutant fiber gene was then combined into a plasmid that contains a full length, E1- and E3-deleted adenovirus genome carrying the above fiber mutation plus a CMV-driven *LacZ* reporter gene in the E1 region. This plasmid was then linearized and transfected into HEK-293 cells expressing the anti-HA pseudo-receptor described in Example 3. After 5 days the cells are freeze-thawed three times, and the virus-containing lysate was passaged onto fresh anti-HA 293 cells.

30 The resultant adenoviruses were further amplified in the anti-HA 293 cells and then purified using standard methods. The vector exhibits reduced binding capacity to CAR on standard HEK-293 cells due to the RLNAEK mutation; however, it binds with high affinity via its HA epitope to the anti-HA pseudoreceptor present on the anti-HA 293 cell line. Moreover, the virus also

35



efficiently infects cells expressing  $\alpha_v$  integrins (such as tumor cells) via the RGD ligand present in the HI loop.

### EXAMPLE 8

5 This example describes an alternatively targeted adenovirus having recombinant fiber proteins exhibiting reduced affinity for the CAR protein and having a non-native ligand.

A mutation was introduced into the Ad2 fiber knob, deleting the Asn-Pro residues in the FG loop (residues 90 and 91 of SEQ ID NO:7). Additionally, the  
10 high-affinity RGD motif was introduced into the HI loop of this protein. The sequences encoding the knob domain were fused to sequences encoding the Ad5 shaft, resulting in a nucleic acid encoding a chimeric Ad5-Ad2 fiber. This construct was cloned into an Ad5 genome also containing the lacZ gene (the AdZ virus), replacing the native fiber sequence. The resultant viruses are termed  
15 AdZ.F\*(RGD).

Increasing particle doses of either AdZ or AdZ.F\*(RGD) were incubated with either SKOV-3 cells (which express both CAR and  $\alpha_v$  integrins) or Ramos cells (which express CAR but not  $\alpha_v$  integrins) in suspension ( $10^6$  cells/300  $\mu$ l medium) for one hour at 36 °C, following which the cells were washed and  
20 incubated overnight. Following the incubation, the cells were assayed for lacZ activity using conventional methods.

The SKOV-3 cells were transduced by both viruses, while the Ramos cells were transduced by AdZ, but only poorly transduced by AdZ.F\*(RGD). These results demonstrate that the native CAR-binding ability of the vector can be  
25 blocked by mutating selective residues of the fiber knob and the virus retarded by the addition of a non-native ligand to the viral coat protein.

### EXAMPLE 9

This example demonstrated the reduced affinity for the CAR protein of  
30 recombinant fiber proteins.

Various cell types (A172, HuVEC, HCAEC, A549, HeLa, HEK-293, and HS68) ( $10^6$  cells/300  $\mu$ l medium) were preincubated for 30 minutes at 37 °C with either soluble Ad5 fiber protein (3 $\mu$ g/ml) or penton base protein (100  $\mu$ g/ml). Following this incubation, either AdZ, AdZ.F\*ab(HA)hi or AdZ.F\*fg(HA)hi (100  
35 viral particles/cell) were added to the cells. After a one hour incubation at 37 °C, the cells were twice washed and incubated overnight, again at 37 °C. Following the incubation, the cells were assayed for lacZ activity using conventional

methods. Except for the HS68 fibroblast cell line, the results indicate that preincubation with Ad5 fiber blocked AdZ transduction, but preincubation with penton base did not. In contrast, the viruses containing the mutant fibers were not blocked by preincubation with fiber, but were blocked by preincubation with penton base. These data are consistent with the ablation of native fiber-based infection through mutating the fibers as indicated.

#### EXAMPLE 10

This example demonstrated the alteration of viral targeting *in vivo*, using an alternatively targeted adenovirus.

The jugular veins of Balb/C mice were injected with either AdZ, AdZ.F\*ab(HA)hi or AdZ.F\*fg(HA)hi ( $10^{10}$  particles/animal in 100  $\mu$ l, eight animals each). The experiments were run in duplicate, and two animals served as a control (100  $\mu$ l saline). At one day post inoculation, the animals were sacrificed and the liver of each was snap-frozen in liquid nitrogen. The livers were then pulverized, and lacZ activity was assayed by conventional methods to determine enzymatic activity/mass of tissue.

The livers from the AdZ.F\*ab(HA)hi- or AdZ.F\*fg(HA)hi-inoculated animals exhibited about 10% of the lacZ activity as those inoculated with AdZ, while control animals exhibited background levels of activity. These results indicate that fiber mutations ablating native cell-receptor binding are effective in greatly reducing native tropism *in vivo*.

All references cited herein are hereby incorporated by reference to the same extent as if each reference were individually and specifically indicated to be incorporated by reference and were set forth in its entirety herein.

While this invention has been described with an emphasis on preferred embodiments, it will be obvious to those of ordinary skill in the art that variations of the preferred embodiments can be used and that it is intended that the invention can be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the invention as defined by the following claims.

**WHAT IS CLAIMED IS:**

1. A recombinant fiber protein comprising an amino terminus of an adenoviral fiber protein and a trimerization domain, wherein said trimerization domain comprises an adenoviral fiber knob domain having a mutation affecting at least one amino acid residue within the region corresponding to the AB loop, B sheet, DE loop, or FG loop of the wild-type Ad5 fiber protein, and wherein said recombinant fiber protein trimerizes when produced in a eukaryotic cell.
2. The recombinant fiber protein of claim 1, wherein said region is the AB loop.
3. The recombinant fiber protein of claim 1, wherein said region is the B sheet.
4. The recombinant fiber protein of claim 1, wherein said amino acid residue corresponds to a residue selected from the group of residues consisting of 408, 409, 412-417, 420, 474-477, and 487-492 of the wild-type Ad5 fiber protein.
5. A recombinant fiber protein comprising an amino terminus of an adenoviral fiber protein and a trimerization domain, wherein said trimerization domain comprises an adenoviral fiber knob having a mutation affecting at least one amino acid corresponding to residue 404-406, 408, 409, 412-417, 420, 439, 441, 442, 449-454, 456, 458, 460, 462, 466, 467, 469-472, 474-477, 482, 485, 487-492, 505-512, 515, 517, 519, 521-528, 533, 535, 537-549, 551, 553, 555, 559-568, 580, or 581 of the wild-type Ad5 fiber protein, and wherein said recombinant fiber protein trimerizes when produced in a eukaryotic cell.
6. The recombinant fiber protein of claim 5, wherein said amino acid residue corresponds to residue 189, 190, 198, 201, or 262 of the native Ad9 fiber protein.
7. The recombinant fiber protein of claim 5, wherein said amino acid residue corresponds to residue 395, 396, 404, 407, or 470 of the native Ad41 long fiber protein.
8. The recombinant fiber protein of claim 5, wherein said amino acid residue corresponds to residue 136, 155, 177, 181, 198, 210, 211, 215, 233, 234, 236, 238, 248, 257, 260, 261, 276, 284, 302, 303, 317, or 318 of the native Ad3 fiber protein.
9. The recombinant fiber protein of any of claims 1-8, wherein said mutation alters the charge of said residue.
10. A trimer comprising the recombinant fiber protein of any of claims 1-8, wherein said trimer has an affinity for a native adenoviral cellular receptor of at least about an order of magnitude less than a wild-type adenoviral fiber trimer.

11. An adenoviral virion comprising the trimer of claim 10.
12. The adenoviral virion of claim 11, comprising a penton base having a mutation affecting at least one native RGD sequence.
13. The adenoviral virion of claim 11, comprising a hexon having a  
5 mutation affecting at least one native HVR sequence.
14. The adenoviral virion of claim 11, lacking a native glycosylation or phosphorylation site.
15. The adenoviral virion of claim 11, which is conjugated to a lipid derivative of polyethylene glycol comprising a primary amine group, an epoxy  
10 group, or a diacylglycerol group.
16. The adenoviral virion of claim 11, which elicits less immunogenicity in a host animal than does a wild-type adenovirus.
17. The adenoviral virion of claim 11, comprising a non-adenoviral ligand.
18. The adenoviral virion of claim 17, wherein said non-adenoviral ligand  
15 is conjugated to a fiber.
19. The adenoviral virion of claim 17, wherein said non-adenoviral ligand is conjugated to a penton.
20. The adenoviral virion of claim 17, wherein said non-adenoviral ligand is conjugated to a hexon.
- 20 21. The adenoviral virion of claim 17, wherein said non-adenoviral ligand is conjugated to protein IX, VI, or IIIa.
22. The adenoviral virion of claim 17, wherein said non-adenoviral ligand binds a substrate other than a native mammalian adenoviral receptor.
23. The adenoviral virion of any of claim 17, wherein said non-adenoviral  
25 ligand binds a substrate other than a native cell-surface protein.
24. The adenoviral virion of claim 17, wherein said substrate is present on the surface of a cell.
25. An adenoviral vector comprising the adenoviral virion of claim 11 and an adenoviral genome.
- 30 26. The adenoviral vector of claim 25, which is replication incompetent.
27. The adenoviral vector of claim 25, which does not productively infect HEK-293 cells.
28. The adenoviral vector of claim 25, wherein said virion comprises a non-adenoviral ligand, and said adenoviral genome comprises a non-native nucleic  
35 acid for transcription.
29. The adenoviral vector of claim 25, wherein said non-native nucleic acid for transcription is operably linked to a non-adenoviral promoter.

30. The adenoviral vector of claim 25, wherein said ligand binds to a substrate present on the surface of a cell and wherein said non-adenoviral promoter is active within said cell.

5 31. The adenoviral vector of claim 29, wherein said non-adenoviral promoter is a tissue-specific promoter.

32. The adenoviral vector of claim 29, wherein said non-adenoviral promoter is a regulable promoter.

33. A method of infecting a cell, comprising contacting a cell with an adenoviral vector of claim 25.

10 34. The method of claim 33, wherein said adenoviral genome comprises a non-native nucleic acid encoding a protein, and wherein said nucleic acid is expressed within said cell to produce said protein.

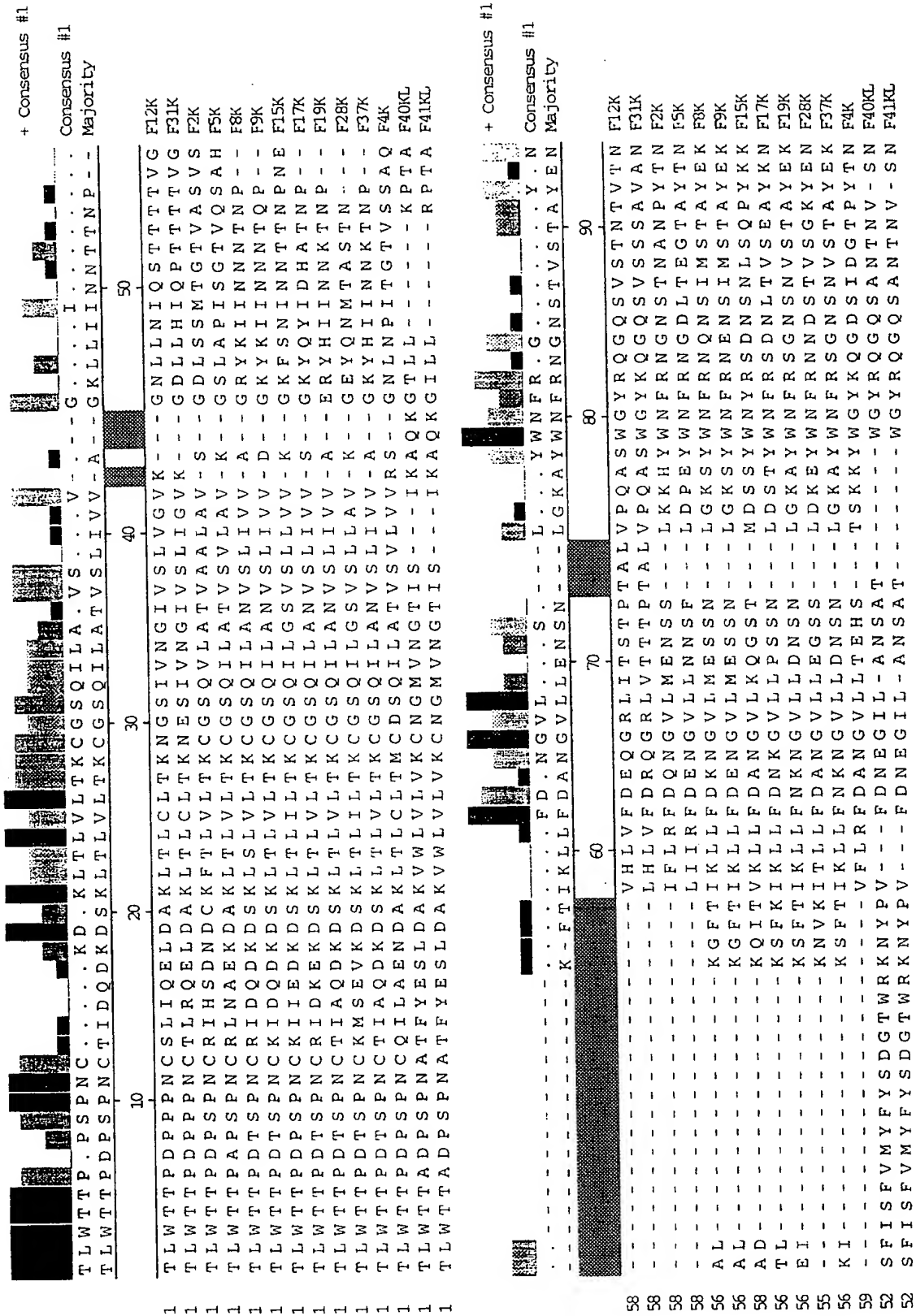
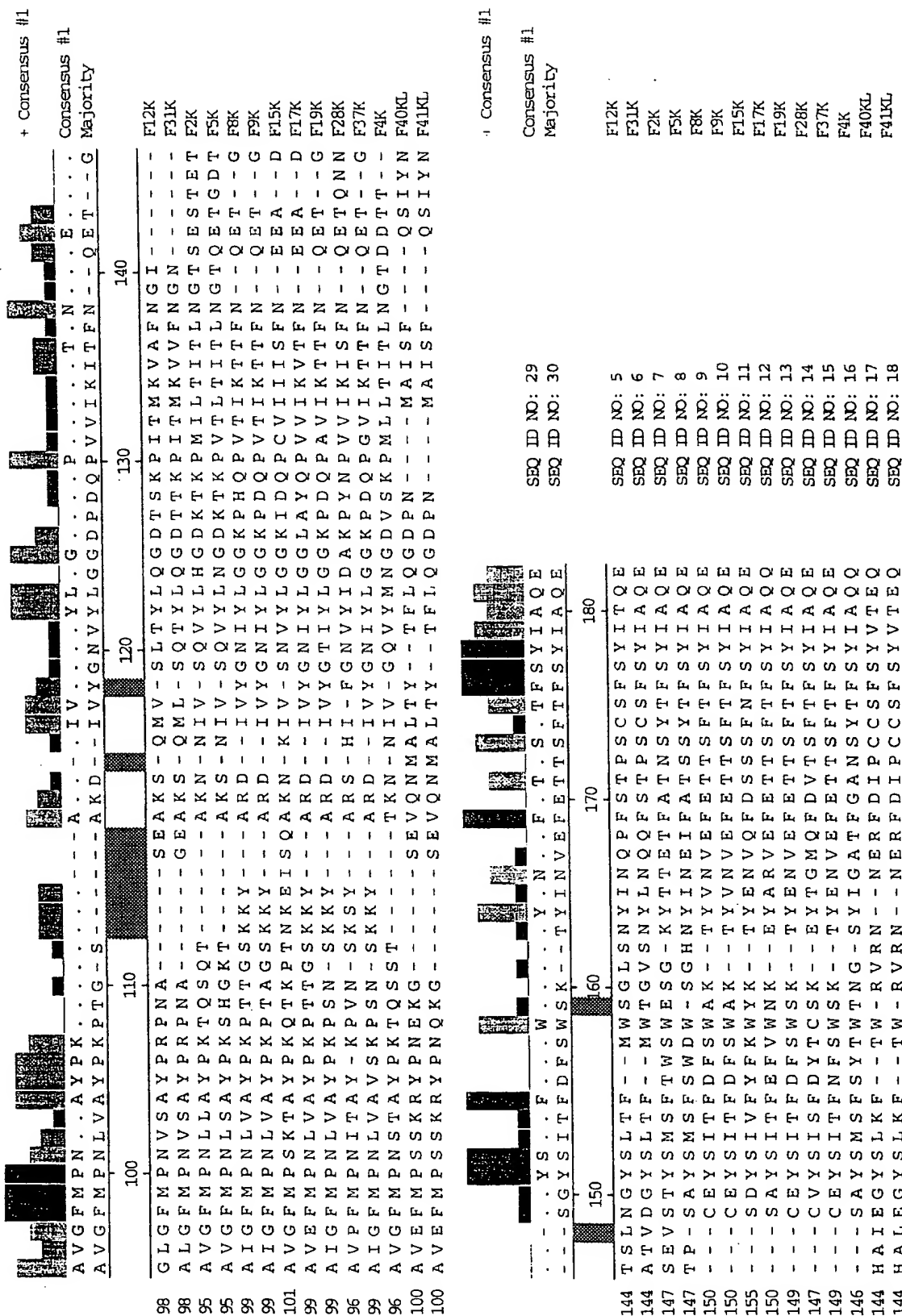
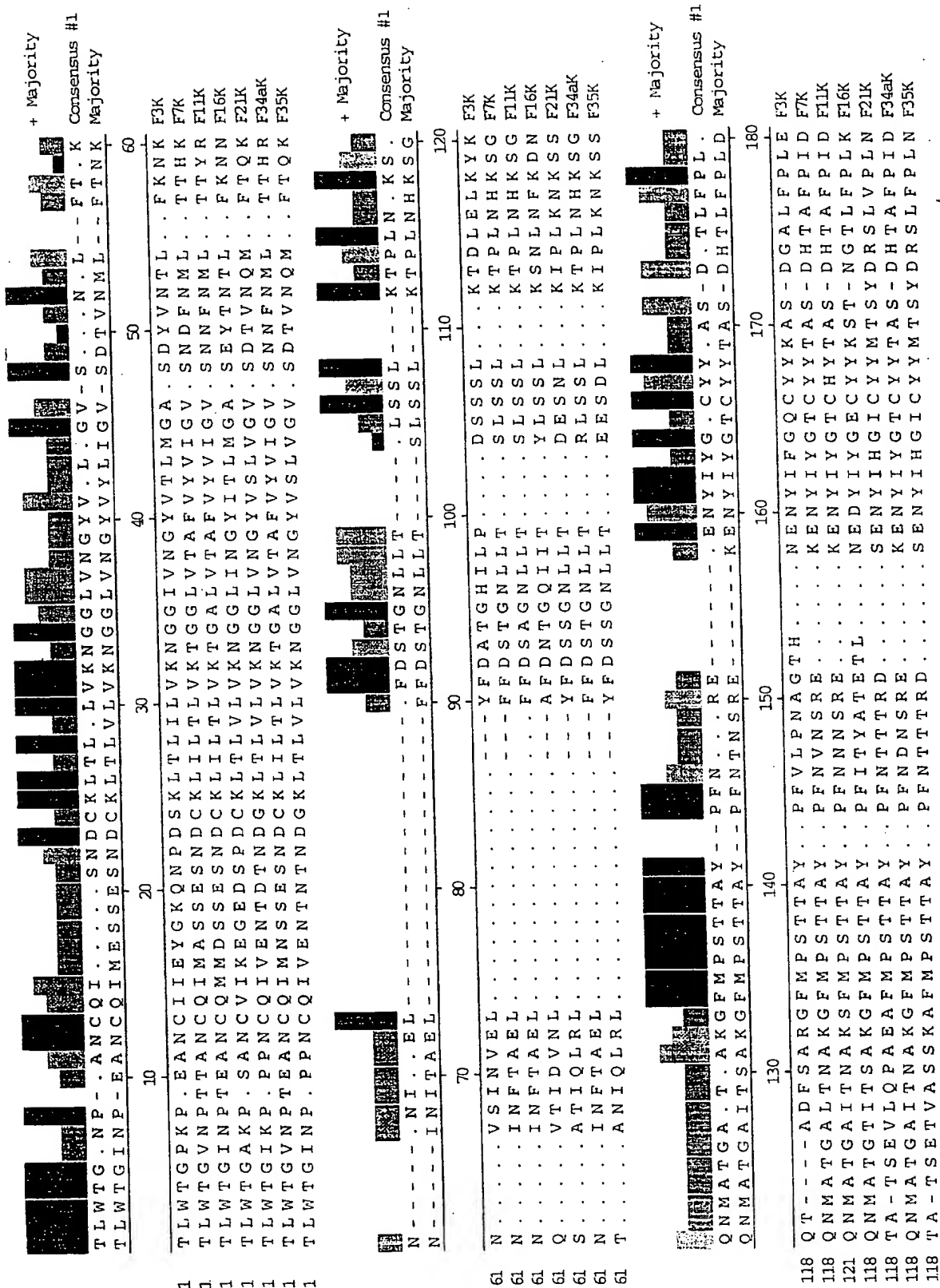


Figure 1A



Consensus 'Consensus #1': When 57% (8) match the residue of F5K, otherwise show '1'.

**Figure 1B**





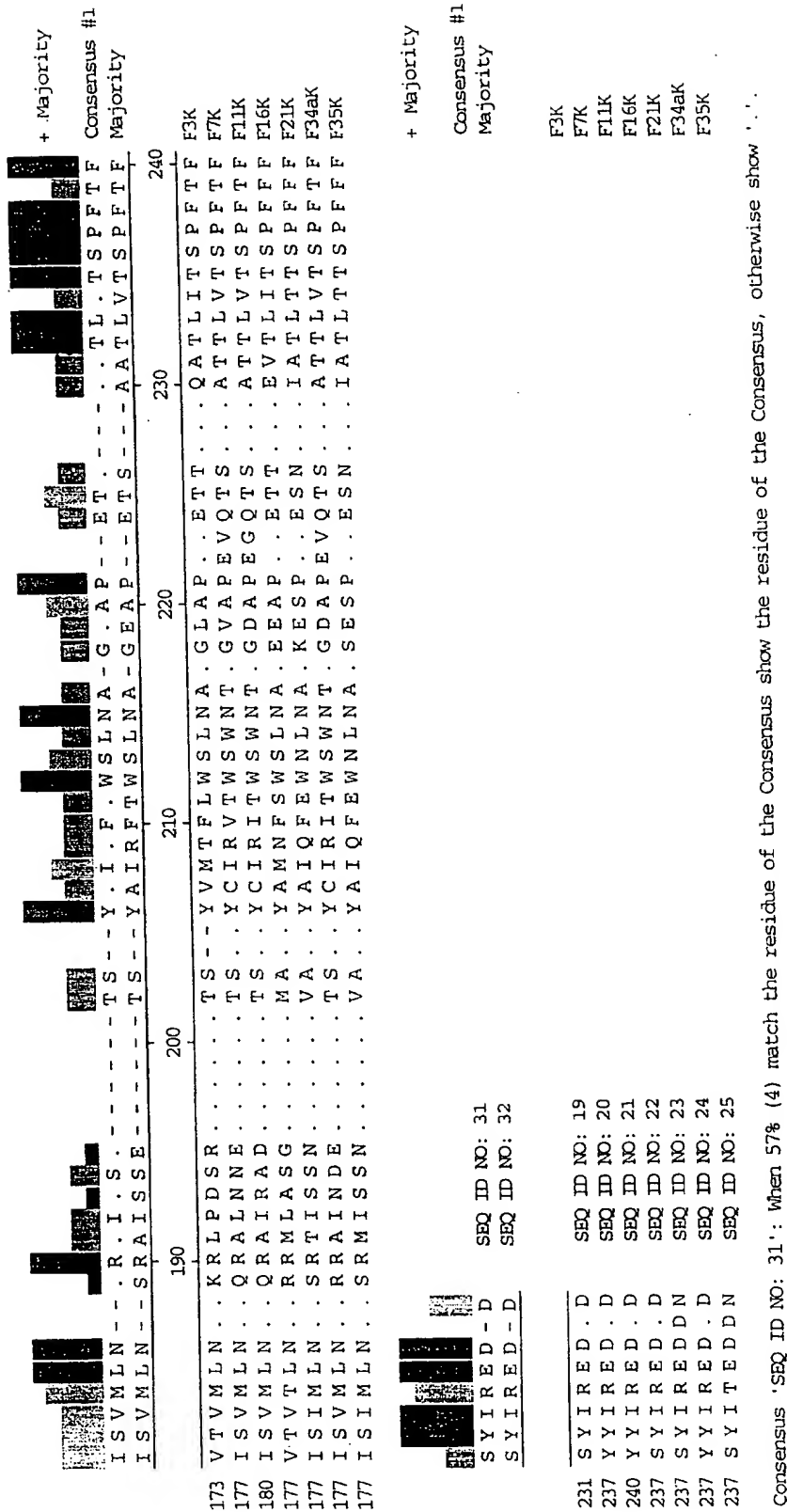


Figure 2B

## SEQUENCE LISTING

&lt;110&gt; GenVec, Inc.

&lt;120&gt; Alternatively Targeted Adenovirus

&lt;130&gt; 202033

&lt;140&gt; WO 99US20728

&lt;141&gt; 199-09-10

&lt;150&gt; US 60/099,851

&lt;151&gt; 1998-09-11

&lt;150&gt; US 60/136,529

&lt;151&gt; 1999-05-28

&lt;160&gt; 32

&lt;170&gt; PatentIn Ver. 2.2

&lt;210&gt; 1

&lt;211&gt; 581

&lt;212&gt; PRT

&lt;213&gt; Human adenovirus serotype 5

&lt;400&gt; 1

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Tyr Asp Thr Glu Thr Gly Pro Pro Thr Val Pro Phe Leu Thr Pro Pro
20      25
Phe Val Ser Pro Asn Gly Phe Gln Glu Ser Pro Pro Gly Val Leu Ser
35      40      45
Leu Arg Leu Ser Glu Pro Leu Val Thr Ser Asn Gly Met Leu Ala Leu
50      55      60
Lys Met Gly Asn Gly Leu Ser Leu Asp Glu Ala Gly Asn Leu Thr Ser
65      70      75      80
Gln Asn Val Thr Thr Val Ser Pro Pro Leu Lys Lys Thr Lys Ser Asn
85      90      95
Ile Asn Leu Glu Ile Ser Ala Pro Leu Thr Val Thr Ser Glu Ala Leu
100     105     110
Thr Val Ala Ala Ala Ala Pro Leu Met Val Ala Gly Asn Thr Leu Thr
115     120     125
Met Gln Ser Gln Ala Pro Leu Thr Val His Asp Ser Lys Leu Ser Ile
130     135     140
Ala Thr Gln Gly Pro Leu Thr Val Ser Glu Gly Lys Leu Ala Leu Gln
145     150     155     160
Thr Ser Gly Pro Leu Thr Thr Thr Asp Ser Ser Thr Leu Thr Ile Thr
165     170     175
Ala Ser Pro Pro Leu Thr Thr Ala Thr Gly Ser Leu Gly Ile Asp Leu
180     185     190
Lys Glu Pro Ile Tyr Thr Gln Asn Gly Lys Leu Gly Leu Lys Tyr Gly
195     200     205
Ala Pro Leu His Val Thr Asp Asp Leu Asn Thr Leu Thr Val Ala Thr
210     215     220
Gly Pro Gly Val Thr Ile Asn Asn Thr Ser Leu Gln Thr Lys Val Thr

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225				230				235				240			
Gly	Ala	Leu	Gly	Phe 245	Asp	Ser	Gln	Gly	Asn 250	Met	Gln	Leu	Asn	Val 255	Ala
Gly	Gly	Leu	Arg 260	Ile	Asp	Ser	Gln	Asn 265	Arg	Arg	Leu	Ile	Leu 270	Asp	Val
Ser	Tyr	Pro 275	Phe	Asp	Ala	Gln	Asn 280	Gln	Leu	Asn	Leu	Arg 285	Leu	Gly	Gln
Gly	Pro 290	Leu	Phe	Ile	Asn	Ser 295	Ala	His	Asn	Leu	Asp 300	Ile	Asn	Tyr	Asn
Lys 305	Gly	Leu	Tyr	Leu	Phe 310	Thr	Ala	Ser	Asn	Asn 315	Ser	Lys	Lys	Leu	Glu 320
Val	Asn	Leu	Ser	Thr 325	Ala	Lys	Gly	Leu	Met 330	Phe	Asp	Ala	Thr	Ala 335	Ile
Ala	Ile	Asn	Ala 340	Gly	Asp	Gly	Leu	Glu 345	Phe	Gly	Ser	Pro	Asn 350	Ala	Pro
Asn	Thr	Asn 355	Pro	Leu	Lys	Thr	Lys 360	Ile	Gly	His	Gly	Leu 365	Glu	Phe	Asp
Ser	Asn 370	Lys	Ala	Met	Val	Pro 375	Lys	Leu	Gly	Thr	Gly 380	Leu	Ser	Phe	Asp
Ser 385	Thr	Gly	Ala	Ile	Thr 390	Val	Gly	Asn	Lys	Asn 395	Asn	Asp	Lys	Leu	Thr 400
Leu	Trp	Thr	Thr	Pro 405	Ala	Pro	Ser	Pro	Asn 410	Cys	Arg	Leu	Asn	Ala 415	Glu
Lys	Asp	Ala	Lys 420	Leu	Thr	Leu	Val	Leu 425	Thr	Lys	Cys	Gly	Ser 430	Gln	Ile
Leu	Ala	Thr 435	Val	Ser	Val	Leu	Ala 440	Val	Lys	Gly	Ser	Leu 445	Ala	Pro	Ile
Ser	Gly 450	Thr	Val	Gln	Ser	Ala 455	His	Leu	Ile	Ile	Arg 460	Phe	Asp	Glu	Asn
Gly 465	Val	Leu	Leu	Asn	Asn 470	Ser	Phe	Leu	Asp	Pro 475	Glu	Tyr	Trp	Asn	Phe 480
Arg	Asn	Gly	Asp	Leu 485	Thr	Glu	Gly	Thr	Ala 490	Tyr	Thr	Asn	Ala	Val 495	Gly
Phe	Met	Pro	Asn 500	Leu	Ser	Ala	Tyr	Pro 505	Lys	Ser	His	Gly	Lys 510	Thr	Ala
Lys	Ser	Asn 515	Ile	Val	Ser	Gln	Val 520	Tyr	Leu	Asn	Gly	Asp 525	Lys	Thr	Lys
Pro	Val 530	Thr	Leu	Thr	Ile	Thr 535	Leu	Asn	Gly	Thr	Gln 540	Glu	Thr	Gly	Asp
Thr 545	Thr	Pro	Ser	Ala	Tyr 550	Ser	Met	Ser	Phe	Ser 555	Trp	Asp	Trp	Ser	Gly 560
His	Asn	Tyr	Ile	Asn 565	Glu	Ile	Phe	Ala	Thr 570	Ser	Ser	Tyr	Thr	Phe 575	Ser
Tyr	Ile	Ala	Gln	Glu											

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<210> 2
<211> 562
<212> PRT
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<213> Human adenovirus serotype 41.LONG

<400> 2

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Met Lys Arg Ala Arg Leu Glu Asp Asp Phe Asn Pro Val Tyr Pro Tyr
1      5      10      15
Glu His Tyr Asn Pro Leu Asp Ile Pro Phe Ile Thr Pro Pro Phe Ala
20      25      30
Ser Ser Asn Gly Leu Gln Glu Lys Pro Pro Gly Val Leu Ser Leu Lys
35      40      45
Tyr Thr Asp Pro Leu Thr Thr Lys Asn Gly Ala Leu Thr Leu Lys Leu
50      55      60
Gly Thr Gly Leu Asn Ile Asp Glu Asn Gly Asp Leu Ser Ser Asp Ala
65      70      75      80
Ser Val Glu Val Ser Ala Pro Ile Thr Lys Thr Asn Lys Ile Val Gly
85      90      95
Leu Asn Tyr Thr Lys Pro Leu Ala Leu Arg Ser Asn Ala Leu Thr Leu
100     105     110
Ser Tyr Asn Ala Pro Leu Asn Val Val Asn Asn Asn Leu Ala Leu Asn
115     120     125
Ile Ser Gln Pro Val Thr Val Asn Ala Asn Asn Glu Leu Ser Leu Leu
130     135     140
Ile Asp Ala Pro Leu Asn Ala Asp Thr Gly Thr Leu Arg Leu Gln Ser
145     150     155     160
Ala Ala Pro Leu Gly Leu Val Asp Lys Thr Leu Lys Val Leu Phe Ser
165     170     175
Ser Pro Leu Tyr Leu Asp Asn Asn Phe Leu Thr Leu Ala Ile Glu Arg
180     185     190
Pro Leu Ala Leu Ser Ser Ser Arg Ala Val Thr Leu Lys Tyr Ser Pro
195     200     205
Pro Leu Lys Ile Glu Asn Glu Asn Leu Thr Leu Ser Thr Gly Gly Pro
210     215     220
Phe Thr Val Ser Gly Gly Asn Leu Asn Leu Thr Thr Ser Ala Pro Leu
225     230     235     240
Ser Val Gln Asn Asn Ser Leu Ser Leu Val Ile Thr Ser Pro Leu Lys
245     250     255
Val Ile Asn Ser Met Leu Ala Val Gly Val Asn Pro Pro Phe Thr Ile
260     265     270
Thr Asp Ser Gly Leu Ala Met Asp Leu Gly Asp Gly Leu Ala Leu Gly
275     280     285
Gly Ser Lys Leu Ile Ile Asn Leu Gly Pro Gly Leu Gln Met Ser Asn
290     295     300
Gly Ala Ile Thr Leu Ala Leu Asp Ala Ala Leu Pro Leu Gln Tyr Arg
305     310     315     320
Asp Asn Gln Leu Gln Leu Arg Ile Gly Ser Thr Ser Gly Leu Ile Met
325     330     335
Ser Gly Val Thr Gln Thr Leu Asn Val Asn Ala Asn Thr Gly Lys Gly
340     345     350
Leu Ala Val Glu Asn Asn Ser Leu Val Val Lys Leu Gly Asn Gly Leu

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355 360 365  
 Arg Phe Asp Ser Trp Gly Ser Ile Thr Val Ser Pro Thr Thr Thr Thr  
 370 375 380  
 Pro Thr Thr Leu Trp Thr Thr Ala Asp Pro Ser Pro Asn Ala Thr Phe  
 385 390 395 400  
 Tyr Glu Ser Leu Asp Ala Lys Val Trp Leu Val Leu Val Lys Cys Asn  
 405 410 415  
 Gly Met Val Asn Gly Thr Ile Ser Ile Lys Ala Gln Lys Gly Ile Leu  
 420 425 430  
 Leu Arg Pro Thr Ala Ser Phe Ile Ser Phe Val Met Tyr Phe Tyr Ser  
 435 440 445  
 Asp Gly Thr Trp Arg Lys Asn Tyr Pro Val Phe Asp Asn Glu Gly Ile  
 450 455 460  
 Leu Ala Asn Ser Ala Thr Trp Gly Tyr Arg Gln Gly Gln Ser Ala Asn  
 465 470 475 480  
 Thr Asn Val Ser Asn Ala Val Glu Phe Met Pro Ser Ser Lys Arg Tyr  
 485 490 495  
 Pro Asn Gln Lys Gly Ser Glu Val Gln Asn Met Ala Leu Thr Tyr Thr  
 500 505 510  
 Phe Leu Gln Gly Asp Pro Asn Met Ala Ile Ser Phe Gln Ser Ile Tyr  
 515 520 525  
 Asn His Ala Leu Glu Gly Tyr Ser Leu Lys Phe Thr Trp Arg Val Arg  
 530 535 540  
 Asn Asn Glu Arg Phe Asp Ile Pro Cys Cys Ser Phe Ser Tyr Val Thr  
 545 550 555 560  
 Glu Gln  
 <210> 3  
 <211> 362  
 <212> PRT  
 <213> Human adenovirus serotype 9  
 <400> 3  
 Met Ser Lys Arg Leu Arg Val Glu Asp Asp Phe Asn Pro Val Tyr Pro  
 1 5 10 15  
 Tyr Gly Tyr Ala Arg Asn Gln Asn Ile Pro Phe Leu Thr Pro Pro Phe  
 20 25 30  
 Val Ser Ser Asp Gly Phe Gln Asn Phe Pro Pro Gly Val Leu Ser Leu  
 35 40 45  
 Lys Leu Ala Asp Pro Ile Ala Ile Val Asn Gly Asn Val Ser Leu Lys  
 50 55 60  
 Val Gly Gly Gly Leu Thr Leu Gln Asp Gly Thr Gly Lys Leu Thr Val  
 65 70 75 80  
 Asn Ala Asp Pro Pro Leu Gln Leu Thr Asn Asn Lys Leu Gly Ile Ala  
 85 90 95  
 Leu Asp Ala Pro Phe Asp Val Ile Asp Asn Lys Leu Thr Leu Leu Ala  
 100 105 110  
 Gly His Gly Leu Ser Ile Ile Thr Lys Glu Thr Ser Thr Leu Pro Gly  
 115 120 125

Leu Arg Asn Thr Leu Val Val Leu Thr Gly Lys Gly Ile Gly Thr Glu  
 130 135 140  
 Ser Thr Asp Asn Gly Gly Thr Val Cys Val Arg Val Gly Glu Gly Gly  
 145 150 155 160  
 Gly Leu Ser Phe Asn Asn Asp Gly Asp Leu Val Ala Phe Asn Lys Lys  
 165 170 175  
 Glu Asp Lys Arg Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys  
 180 185 190  
 Lys Ile Asp Gln Asp Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys  
 195 200 205  
 Cys Gly Ser Gln Ile Leu Ala Asn Val Ser Leu Ile Val Val Asp Gly  
 210 215 220  
 Lys Tyr Lys Ile Ile Asn Asn Asn Thr Gln Pro Ala Leu Lys Gly Phe  
 225 230 235 240  
 Thr Ile Lys Leu Leu Phe Asp Glu Asn Gly Val Leu Met Glu Ser Ser  
 245 250 255  
 Asn Leu Gly Lys Ser Tyr Trp Asn Phe Arg Asn Glu Asn Ser Ile Met  
 260 265 270  
 Ser Thr Ala Tyr Glu Lys Ala Ile Gly Phe Met Pro Asn Leu Val Ala  
 275 280 285  
 Tyr Pro Lys Pro Thr Ala Gly Ser Lys Lys Tyr Ala Arg Asp Ile Val  
 290 295 300  
 Tyr Gly Asn Ile Tyr Leu Gly Gly Lys Pro Asp Gln Pro Val Thr Ile  
 305 310 315 320  
 Lys Thr Thr Phe Asn Gln Glu Thr Gly Cys Glu Tyr Ser Ile Thr Phe  
 325 330 335  
 Asp Phe Ser Trp Ala Lys Thr Tyr Val Asn Val Glu Phe Glu Thr Thr  
 340 345 350  
 Ser Phe Thr Phe Ser Tyr Ile Ala Gln Glu  
 355 360

<210> 4  
 <211> 319  
 <212> PRT  
 <213> Human adenovirus serotype 3

<400> 4

Met Ala Lys Arg Ala Arg Leu Ser Thr Ser Phe Asn Pro Val Tyr Pro  
 1 5 10 15  
 Tyr Glu Asp Glu Ser Ser Ser Gln His Pro Phe Ile Asn Pro Gly Phe  
 20 25 30  
 Ile Ser Pro Asp Gly Phe Thr Gln Ser Pro Asn Gly Val Leu Ser Leu  
 35 40 45  
 Lys Cys Val Asn Pro Leu Thr Thr Ala Ser Gly Ser Leu Gln Leu Lys  
 50 55 60  
 Val Gly Ser Gly Leu Thr Val Asp Thr Thr Asp Gly Ser Leu Glu Glu  
 65 70 75 80  
 Asn Ile Lys Val Asn Thr Pro Leu Thr Lys Ser Asn His Ser Ile Asn  
 85 90 95  
 Leu Pro Ile Gly Asn Gly Leu Gln Ile Glu Gln Asn Lys Leu Cys Ser

100 105 110  
 Lys Leu Gly Asn Gly Leu Thr Phe Asp Ser Ser Asn Ser Ile Ala Leu  
 115 120 125  
 Lys Asn Asn Thr Leu Trp Thr Gly Pro Lys Pro Glu Ala Asn Cys Ile  
 130 135 140  
 Ile Glu Tyr Gly Lys Gln Asn Pro Asp Ser Lys Leu Thr Leu Ile Leu  
 145 150 155 160  
 Val Lys Asn Gly Gly Ile Val Asn Gly Tyr Val Thr Leu Met Gly Ala  
 165 170 175  
 Ser Asp Tyr Val Asn Thr Leu Phe Lys Asn Lys Asn Val Ser Ile Asn  
 180 185 190  
 Val Glu Leu Tyr Phe Asp Ala Thr Gly His Ile Leu Pro Asp Ser Ser  
 195 200 205  
 Ser Leu Lys Thr Asp Leu Glu Leu Lys Tyr Lys Gln Thr Ala Asp Phe  
 210 215 220  
 Ser Ala Arg Gly Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe Val Leu  
 225 230 235 240  
 Pro Asn Ala Gly Thr His Asn Glu Asn Tyr Ile Phe Gly Gln Cys Tyr  
 245 250 255  
 Tyr Lys Ala Ser Asp Gly Ala Leu Phe Pro Leu Glu Val Thr Val Met  
 260 265 270  
 Leu Asn Lys Arg Leu Pro Asp Ser Arg Thr Ser Tyr Val Met Thr Phe  
 275 280 285  
 Leu Trp Ser Leu Asn Ala Gly Leu Ala Pro Glu Thr Thr Gln Ala Thr  
 290 295 300  
 Leu Ile Thr Ser Pro Phe Thr Phe Ser Tyr Ile Arg Glu Asp Asp  
 305 310 315  
 <210> 5  
 <211> 179  
 <212> PRT  
 <213> Human adenovirus serotype 12  
 <400> 5  
 Thr Leu Trp Thr Thr Pro Asp Pro Pro Pro Asn Cys Ser Leu Ile Gln  
 1 5 10 15  
 Glu Leu Asp Ala Lys Leu Thr Leu Cys Leu Thr Lys Asn Gly Ser Ile  
 20 25 30  
 Val Asn Gly Ile Val Ser Leu Val Gly Val Lys Gly Asn Leu Leu Asn  
 35 40 45  
 Ile Gln Ser Thr Thr Thr Thr Val Gly Val His Leu Val Phe Asp Glu  
 50 55 60  
 Gln Gly Arg Leu Ile Thr Ser Thr Pro Thr Ala Leu Val Pro Gln Ala  
 65 70 75 80  
 Ser Trp Gly Tyr Arg Gln Gly Gln Ser Val Ser Thr Asn Thr Val Thr  
 85 90 95  
 Asn Gly Leu Gly Phe Met Pro Asn Val Ser Ala Tyr Pro Arg Pro Asn  
 100 105 110  
 Ala Ser Glu Ala Lys Ser Gln Met Val Ser Leu Thr Tyr Leu Gln Gly  
 115 120 125

Asp Thr Ser Lys Pro Ile Thr Met Lys Val Ala Phe Asn Gly Ile Thr  
 130 135 140  
 Ser Leu Asn Gly Tyr Ser Leu Thr Phe Met Trp Ser Gly Leu Ser Asn  
 145 150 155 160  
 Tyr Ile Asn Gln Pro Phe Ser Thr Pro Ser Cys Ser Phe Ser Tyr Ile  
 165 170 175

Thr Gln Glu

<210> 6  
 <211> 179  
 <212> PRT  
 <213> Human adenovirus serotype 31

<400> 6

Thr Leu Trp Thr Thr Pro Asp Pro Pro Pro Asn Cys Thr Leu Arg Gln  
 1 5 10 15  
 Glu Leu Asp Ala Lys Leu Thr Leu Cys Leu Thr Lys Asn Glu Ser Ile  
 20 25 30  
 Val Asn Gly Ile Val Ser Leu Ile Gly Val Lys Gly Asp Leu Leu His  
 35 40 45  
 Ile Gln Pro Thr Thr Thr Thr Val Gly Leu His Leu Val Phe Asp Arg  
 50 55 60  
 Gln Gly Arg Leu Val Thr Thr Thr Pro Thr Ala Leu Val Pro Gln Ala  
 65 70 75 80  
 Ser Trp Gly Tyr Lys Gln Gly Gln Ser Val Ser Ser Ser Ala Val Ala  
 85 90 95  
 Asn Ala Leu Gly Phe Met Pro Asn Val Ser Ala Tyr Pro Arg Pro Asn  
 100 105 110  
 Ala Gly Glu Ala Lys Ser Gln Met Leu Ser Gln Thr Tyr Leu Gln Gly  
 115 120 125  
 Asp Thr Thr Lys Pro Ile Thr Met Lys Val Val Phe Asn Gly Asn Ala  
 130 135 140  
 Thr Val Asp Gly Tyr Ser Leu Thr Phe Met Trp Thr Gly Val Ser Asn  
 145 150 155 160  
 Tyr Leu Asn Gln Gln Phe Ser Thr Pro Ser Cys Ser Phe Ser Tyr Ile  
 165 170 175

Ala Gln Glu

<210> 7  
 <211> 183  
 <212> PRT  
 <213> Human adenovirus serotype 2

<400> 7

Thr Leu Trp Thr Thr Pro Asp Pro Ser Pro Asn Cys Arg Ile His Ser  
 1 5 10 15  
 Asp Asn Asp Cys Lys Phe Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Val Leu Ala Thr Val Ala Ala Leu Ala Val Ser Gly Asp Leu Ser Ser  
 35 40 45  
 Met Thr Gly Thr Val Ala Ser Val Ser Ile Phe Leu Arg Phe Asp Gln



8

50                      55                      60  
 Asn Gly Val Leu Met Glu Asn Ser Ser Leu Lys Lys His Tyr Trp Asn  
 65                      70                      75                      80  
 Phe Arg Asn Gly Asn Ser Thr Asn Ala Asn Pro Tyr Thr Asn Ala Val  
                     85                      90                      95  
 Gly Phe Met Pro Asn Leu Leu Ala Tyr Pro Lys Thr Gln Ser Gln Thr  
                     100                      105                      110  
 Ala Lys Asn Asn Ile Val Ser Gln Val Tyr Leu His Gly Asp Lys Thr  
                     115                      120                      125  
 Lys Pro Met Ile Leu Thr Ile Thr Leu Asn Gly Thr Ser Glu Ser Thr  
                     130                      135                      140  
 Glu Thr Ser Glu Val Ser Thr Tyr Ser Met Ser Phe Thr Trp Ser Trp  
 145                      150                      155                      160  
 Glu Ser Gly Lys Tyr Thr Thr Glu Thr Phe Ala Thr Asn Ser Tyr Thr  
                     165                      170                      175  
 Phe Ser Tyr Ile Ala Gln Glu  
                     180

<210> 8  
 <211> 182  
 <212> PRT  
 <213> Human adenovirus serotype 5

<400> 8

Thr Leu Trp Thr Thr Pro Ala Pro Ser Pro Asn Cys Arg Leu Asn Ala  
 1                      5                      10                      15  
 Glu Lys Asp Ala Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
                     20                      25                      30  
 Ile Leu Ala Thr Val Ser Val Leu Ala Val Lys Gly Ser Leu Ala Pro  
                     35                      40                      45  
 Ile Ser Gly Thr Val Gln Ser Ala His Leu Ile Ile Arg Phe Asp Glu  
                     50                      55                      60  
 Asn Gly Val Leu Leu Asn Asn Ser Phe Leu Asp Pro Glu Tyr Trp Asn  
 65                      70                      75                      80  
 Phe Arg Asn Gly Asp Leu Thr Glu Gly Thr Ala Tyr Thr Asn Ala Val  
                     85                      90                      95  
 Gly Phe Met Pro Asn Leu Ser Ala Tyr Pro Lys Ser His Gly Lys Thr  
                     100                      105                      110  
 Ala Lys Ser Asn Ile Val Ser Gln Val Tyr Leu Asn Gly Asp Lys Thr  
                     115                      120                      125  
 Lys Pro Val Thr Leu Thr Ile Thr Leu Asn Gly Thr Gln Glu Thr Gly  
                     130                      135                      140  
 Asp Thr Thr Pro Ser Ala Tyr Ser Met Ser Phe Ser Trp Asp Trp Ser  
 145                      150                      155                      160  
 Gly His Asn Tyr Ile Asn Glu Ile Phe Ala Thr Ser Ser Tyr Thr Phe  
                     165                      170                      175  
 Ser Tyr Ile Ala Gln Glu  
                     180

<210> 9  
 <211> 182

<212> PRT  
 <213> Human adenovirus serotype 8

<400> 9

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Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Arg Ile Asp Gln
1      5      10      15
Asp Lys Asp Ser Lys Leu Ser Leu Val Leu Thr Lys Cys Gly Ser Gln
20      25      30
Ile Leu Ala Asn Val Ser Leu Ile Val Val Ala Gly Arg Tyr Lys Ile
35      40      45
Ile Asn Asn Asn Thr Asn Pro Ala Leu Lys Gly Phe Thr Ile Lys Leu
50      55      60
Leu Phe Asp Lys Asn Gly Val Leu Met Glu Ser Ser Asn Leu Gly Lys
65      70      75      80
Ser Tyr Trp Asn Phe Arg Asn Gln Asn Ser Ile Met Ser Thr Ala Tyr
85      90      95
Glu Lys Ala Ile Gly Phe Met Pro Asn Leu Val Ala Tyr Pro Lys Pro
100     105     110
Thr Thr Gly Ser Lys Lys Tyr Ala Arg Asp Ile Val Tyr Gly Asn Ile
115     120     125
Tyr Leu Gly Gly Lys Pro His Gln Pro Val Thr Ile Lys Thr Thr Phe
130     135     140
Asn Gln Glu Thr Gly Cys Glu Tyr Ser Ile Thr Phe Asp Phe Ser Trp
145     150     155     160
Ala Lys Thr Tyr Val Asn Val Glu Phe Glu Thr Thr Ser Phe Thr Phe
165     170     175
Ser Tyr Ile Ala Gln Glu
180

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<210> 10  
 <211> 182  
 <212> PRT  
 <213> Human adenovirus serotype 9

<400> 10

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Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Lys Ile Asp Gln
1      5      10      15
Asp Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln
20      25      30
Ile Leu Ala Asn Val Ser Leu Ile Val Val Asp Gly Lys Tyr Lys Ile
35      40      45
Ile Asn Asn Asn Thr Gln Pro Ala Leu Lys Gly Phe Thr Ile Lys Leu
50      55      60
Leu Phe Asp Glu Asn Gly Val Leu Met Glu Ser Ser Asn Leu Gly Lys
65      70      75      80
Ser Tyr Trp Asn Phe Arg Asn Glu Asn Ser Ile Met Ser Thr Ala Tyr
85      90      95
Glu Lys Ala Ile Gly Phe Met Pro Asn Leu Val Ala Tyr Pro Lys Pro
100     105     110
Thr Ala Gly Ser Lys Lys Tyr Ala Arg Asp Ile Val Tyr Gly Asn Ile
115     120     125

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10

Tyr Leu Gly Gly Lys Pro Asp Gln Pro Val Thr Ile Lys Thr Thr Phe  
 130 135 140  
 Asn Gln Glu Thr Gly Cys Glu Tyr Ser Ile Thr Phe Asp Phe Ser Trp  
 145 150 155 160  
 Ala Lys Thr Tyr Val Asn Val Glu Phe Glu Thr Thr Ser Phe Thr Phe  
 165 170 175  
 Ser Tyr Ile Ala Gln Glu  
 180

<210> 11  
 <211> 187  
 <212> PRT  
 <213> Human adenovirus serotype 15

<400> 11

Thr Leu Trp Thr Thr Pro Asp Pro Ser Pro Asn Cys Lys Ile Ile Glu  
 1 5 10 15  
 Asp Lys Asp Ser Lys Leu Thr Leu Ile Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Gly Ser Val Ser Leu Leu Val Val Lys Gly Lys Phe Ser Asn  
 35 40 45  
 Ile Asn Asn Thr Thr Asn Pro Asn Glu Ala Asp Lys Gln Ile Thr Val  
 50 55 60  
 Lys Leu Leu Phe Asp Ala Asn Gly Val Leu Lys Gln Gly Ser Thr Met  
 65 70 75 80  
 Asp Ser Ser Tyr Trp Asn Tyr Arg Ser Asp Asn Ser Asn Leu Ser Gln  
 85 90 95  
 Pro Tyr Lys Lys Ala Val Gly Phe Met Pro Ser Lys Thr Ala Tyr Pro  
 100 105 110  
 Lys Gln Thr Lys Pro Thr Asn Lys Glu Ile Ser Gln Ala Lys Asn Lys  
 115 120 125  
 Ile Val Ser Asn Val Tyr Leu Gly Gly Lys Ile Asp Gln Pro Cys Val  
 130 135 140  
 Ile Ile Ile Ser Phe Asn Glu Glu Ala Asp Ser Asp Tyr Ser Ile Val  
 145 150 155 160  
 Phe Tyr Phe Lys Trp Tyr Lys Thr Tyr Glu Asn Val Gln Phe Asp Ser  
 165 170 175  
 Ser Ser Phe Asn Phe Ser Tyr Ile Ala Gln Glu  
 180 185

<210> 12  
 <211> 182  
 <212> PRT  
 <213> Human adenovirus serotype 17

<400> 12

Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Arg Ile Asp Lys  
 1 5 10 15  
 Glu Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Ala Asn Val Ser Leu Ile Val Val Ser Gly Lys Tyr Gln Tyr  
 35 40 45

11

Ile Asp His Ala Thr Asn Pro Thr Leu Lys Ser Phe Lys Ile Lys Leu  
 50 55 60  
 Leu Phe Asp Asn Lys Gly Val Leu Leu Pro Ser Ser Asn Leu Asp Ser  
 65 70 75 80  
 Thr Tyr Trp Asn Phe Arg Ser Asp Asn Leu Thr Val Ser Glu Ala Tyr  
 85 90 95  
 Lys Asn Ala Val Glu Phe Met Pro Asn Leu Val Ala Tyr Pro Lys Pro  
 100 105 110  
 Thr Thr Gly Ser Lys Lys Tyr Ala Arg Asp Ile Val Tyr Gly Asn Ile  
 115 120 125  
 Tyr Leu Gly Gly Leu Ala Tyr Gln Pro Val Val Ile Lys Val Thr Phe  
 130 135 140  
 Asn Glu Glu Ala Asp Ser Ala Tyr Ser Ile Thr Phe Glu Phe Val Trp  
 145 150 155 160  
 Asn Lys Glu Tyr Ala Arg Val Glu Phe Glu Thr Thr Ser Phe Thr Phe  
 165 170 175  
 Ser Tyr Ile Ala Gln Gln  
 180

<210> 13  
 <211> 181  
 <212> PRT  
 <213> Human adenovirus serotype 19

<400> 13

Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Thr Ile Ala Gln  
 1 5 10 15  
 Asp Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Ala Asn Val Ser Leu Ile Val Val Ala Glu Arg Tyr His Ile  
 35 40 45  
 Ile Asn Asn Lys Thr Asn Pro Glu Ile Lys Ser Phe Thr Ile Lys Leu  
 50 55 60  
 Leu Phe Asn Lys Asn Gly Val Leu Leu Asp Asn Ser Asn Leu Gly Lys  
 65 70 75 80  
 Ala Tyr Trp Asn Phe Arg Ser Gly Asn Ser Asn Val Ser Thr Ala Tyr  
 85 90 95  
 Glu Lys Ala Ile Gly Phe Met Pro Asn Leu Val Ala Tyr Pro Lys Pro  
 100 105 110  
 Ser Asn Ser Lys Lys Tyr Ala Arg Asp Ile Val Tyr Gly Thr Ile Tyr  
 115 120 125  
 Leu Gly Gly Lys Pro Asp Gln Pro Ala Val Ile Lys Thr Thr Phe Asn  
 130 135 140  
 Gln Glu Thr Gly Cys Glu Tyr Ser Ile Thr Phe Asp Phe Ser Trp Ser  
 145 150 155 160  
 Lys Thr Tyr Glu Asn Val Glu Phe Glu Thr Thr Ser Phe Thr Phe Ser  
 165 170 175  
 Tyr Ile Ala Gln Glu  
 180

12

<210> 14  
 <211> 179  
 <212> PRT  
 <213> Human adenovirus serotype 28

<400> 14

Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Lys Met Ser Glu  
 1 5 10 15  
 Val Lys Asp Ser Lys Leu Thr Leu Ile Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Gly Ser Val Ser Leu Leu Ala Val Lys Gly Glu Tyr Gln Asn  
 35 40 45  
 Met Thr Ala Ser Thr Asn Lys Asn Val Lys Ile Thr Leu Leu Phe Asp  
 50 55 60  
 Ala Asn Gly Val Leu Leu Glu Gly Ser Ser Leu Asp Lys Glu Tyr Trp  
 65 70 75 80  
 Asn Phe Arg Asn Asn Asp Ser Thr Val Ser Gly Lys Tyr Glu Asn Ala  
 85 90 95  
 Val Pro Phe Met Pro Asn Ile Thr Ala Tyr Lys Pro Val Asn Ser Lys  
 100 105 110  
 Ser Tyr Ala Arg Ser His Ile Phe Gly Asn Val Tyr Ile Asp Ala Lys  
 115 120 125  
 Pro Tyr Asn Pro Val Val Ile Lys Ile Ser Phe Asn Gln Glu Thr Gln  
 130 135 140  
 Asn Asn Cys Val Tyr Ser Ile Ser Phe Asp Tyr Thr Cys Ser Lys Glu  
 145 150 155 160  
 Tyr Thr Gly Met Gln Phe Asp Val Thr Ser Phe Thr Phe Ser Tyr Ile  
 165 170 175  
 Ala Gln Glu

<210> 15  
 <211> 181  
 <212> PRT  
 <213> Human adenovirus serotype 37

<400> 15

Thr Leu Trp Thr Thr Pro Asp Thr Ser Pro Asn Cys Thr Ile Ala Gln  
 1 5 10 15  
 Asp Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Ala Asn Val Ser Leu Ile Val Val Ala Gly Lys Tyr His Ile  
 35 40 45  
 Ile Asn Asn Lys Thr Asn Pro Lys Ile Lys Ser Phe Thr Ile Lys Leu  
 50 55 60  
 Leu Phe Asn Lys Asn Gly Val Leu Leu Asp Asn Ser Asn Leu Gly Lys  
 65 70 75 80  
 Ala Tyr Trp Asn Phe Arg Ser Gly Asn Ser Asn Val Ser Thr Ala Tyr  
 85 90 95  
 Glu Lys Ala Ile Gly Phe Met Pro Asn Leu Val Ala Val Ser Lys Pro  
 100 105 110  
 Ser Asn Ser Lys Lys Tyr Ala Arg Asp Ile Val Tyr Gly Asn Ile Tyr

13

115                      120                      125  
 Leu Gly Gly Lys Pro Asp Gln Pro Gly Val Ile Lys Thr Thr Phe Asn  
     130                      135                      140  
 Gln Glu Thr Gly Cys Glu Tyr Ser Ile Thr Phe Asn Phe Ser Trp Ser  
     145                      150                      155                      160  
 Lys Thr Tyr Glu Asn Val Glu Phe Glu Thr Thr Ser Phe Thr Phe Ser  
                     165                      170                      175  
 Tyr Ile Ala Gln Glu  
                     180

<210> 16  
 <211> 179  
 <212> PRT  
 <213> Human adenovirus serotype 4

<400> 16

Thr Leu Trp Thr Thr Pro Asp Pro Ser Pro Asn Cys Gln Ile Leu Ala  
     1                      5                      10                      15  
 Glu Asn Asp Ala Lys Leu Thr Leu Cys Leu Thr Met Cys Asp Ser Gln  
                     20                      25                      30  
 Ile Leu Ala Thr Val Ser Val Leu Val Val Arg Ser Gly Asn Leu Asn  
                     35                      40                      45  
 Pro Ile Thr Gly Thr Val Ser Ser Ala Gln Val Phe Leu Arg Phe Asp  
     50                      55                      60  
 Ala Asn Gly Val Leu Leu Thr Glu His Ser Thr Ser Lys Lys Tyr Trp  
     65                      70                      75                      80  
 Gly Tyr Lys Gln Gly Asp Ser Ile Asp Gly Thr Pro Tyr Thr Asn Ala  
                     85                      90                      95  
 Val Gly Phe Met Pro Asn Ser Thr Ala Tyr Pro Lys Thr Gln Ser Ser  
                     100                      105                      110  
 Thr Thr Lys Asn Asn Ile Val Gly Gln Val Tyr Met Asn Gly Asp Val  
                     115                      120                      125  
 Ser Lys Pro Met Leu Leu Thr Ile Thr Leu Asn Gly Thr Asp Asp Thr  
     130                      135                      140  
 Thr Ser Ala Tyr Ser Met Ser Phe Ser Tyr Thr Trp Thr Asn Gly Ser  
     145                      150                      155                      160  
 Tyr Ile Gly Ala Thr Phe Gly Ala Asn Ser Tyr Thr Phe Ser Tyr Ile  
                     165                      170                      175

Ala Gln Gln

<210> 17  
 <211> 176  
 <212> PRT  
 <213> Human adenovirus serotype 40LONG

<400> 17

Thr Leu Trp Thr Thr Ala Asp Pro Ser Pro Asn Ala Thr Phe Tyr Glu  
     1                      5                      10                      15  
 Ser Leu Asp Ala Lys Val Trp Leu Val Leu Val Lys Cys Asn Gly Met  
                     20                      25                      30  
 Val Asn Gly Thr Ile Ser Ile Lys Ala Gln Lys Gly Thr Leu Leu Lys  
     35                      40                      45

14

Pro Thr Ala Ser Phe Ile Ser Phe Val Met Tyr Phe Tyr Ser Asp Gly  
 50 55 60  
 Thr Trp Arg Lys Asn Tyr Pro Val Phe Asp Asn Glu Gly Ile Leu Ala  
 65 70 75 80  
 Asn Ser Ala Thr Trp Gly Tyr Arg Gln Gly Gln Ser Ala Asn Thr Asn  
 85 90 95  
 Val Ser Asn Ala Val Glu Phe Met Pro Ser Ser Lys Arg Tyr Pro Asn  
 100 105 110  
 Glu Lys Gly Ser Glu Val Gln Asn Met Ala Leu Thr Tyr Thr Phe Leu  
 115 120 125  
 Gln Gly Asp Pro Asn Met Ala Ile Ser Phe Gln Ser Ile Tyr Asn His  
 130 135 140  
 Ala Ile Glu Gly Tyr Ser Leu Lys Phe Thr Trp Arg Val Arg Asn Asn  
 145 150 155 160  
 Glu Arg Phe Asp Ile Pro Cys Cys Ser Phe Ser Tyr Val Thr Glu Gln  
 165 170 175

<210> 18  
 <211> 176  
 <212> PRT  
 <213> Human adenovirus serotype 41LONG

<400> 18

Thr Leu Trp Thr Thr Ala Asp Pro Ser Pro Asn Ala Thr Phe Tyr Glu  
 1 5 10 15  
 Ser Leu Asp Ala Lys Val Trp Leu Val Leu Val Lys Cys Asn Gly Met  
 20 25 30  
 Val Asn Gly Thr Ile Ser Ile Lys Ala Gln Lys Gly Ile Leu Leu Arg  
 35 40 45  
 Pro Thr Ala Ser Phe Ile Ser Phe Val Met Tyr Phe Tyr Ser Asp Gly  
 50 55 60  
 Thr Trp Arg Lys Asn Tyr Pro Val Phe Asp Asn Glu Gly Ile Leu Ala  
 65 70 75 80  
 Asn Ser Ala Thr Trp Gly Tyr Arg Gln Gly Gln Ser Ala Asn Thr Asn  
 85 90 95  
 Val Ser Asn Ala Val Glu Phe Met Pro Ser Ser Lys Arg Tyr Pro Asn  
 100 105 110  
 Gln Lys Gly Ser Glu Val Gln Asn Met Ala Leu Thr Tyr Thr Phe Leu  
 115 120 125  
 Gln Gly Asp Pro Asn Met Ala Ile Ser Phe Gln Ser Ile Tyr Asn His  
 130 135 140  
 Ala Leu Glu Gly Tyr Ser Leu Lys Phe Thr Trp Arg Val Arg Asn Asn  
 145 150 155 160  
 Glu Arg Phe Asp Ile Pro Cys Cys Ser Phe Ser Tyr Val Thr Glu Gln  
 165 170 175

<210> 19  
 <211> 188  
 <212> PRT  
 <213> Human adenovirus serotype 3

<400> 19

15

Thr Leu Trp Thr Gly Pro Lys Pro Glu Ala Asn Cys Ile Ile Glu Tyr  
 1 5 10 15  
 Gly Lys Gln Asn Pro Asp Ser Lys Leu Thr Leu Ile Leu Val Lys Asn  
 20 25 30  
 Gly Gly Ile Val Asn Gly Tyr Val Thr Leu Met Gly Ala Ser Asp Tyr  
 35 40 45  
 Val Asn Thr Leu Phe Lys Asn Lys Asn Val Ser Ile Asn Val Glu Leu  
 50 55 60  
 Tyr Phe Asp Ala Thr Gly His Ile Leu Pro Asp Ser Ser Ser Leu Lys  
 65 70 75 80  
 Thr Asp Leu Glu Leu Lys Tyr Lys Gln Thr Ala Asp Phe Ser Ala Arg  
 85 90 95  
 Gly Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe Val Leu Pro Asn Ala  
 100 105 110  
 Gly Thr His Asn Glu Asn Tyr Ile Phe Gly Gln Cys Tyr Tyr Lys Ala  
 115 120 125  
 Ser Asp Gly Ala Leu Phe Pro Leu Glu Val Thr Val Met Leu Asn Lys  
 130 135 140  
 Arg Leu Pro Asp Ser Arg Thr Ser Tyr Val Met Thr Phe Leu Trp Ser  
 145 150 155 160  
 Leu Asn Ala Gly Leu Ala Pro Glu Thr Thr Gln Ala Thr Leu Ile Thr  
 165 170 175  
 Ser Pro Phe Thr Phe Ser Tyr Ile Arg Glu Asp Asp  
 180 185

<210> 20  
 <211> 193  
 <212> PRT  
 <213> Human adenovirus serotype 7

<400> 20

Thr Leu Trp Thr Gly Val Asn Pro Thr Thr Ala Asn Cys Gln Ile Met  
 1 5 10 15  
 Ala Ser Ser Glu Ser Asn Asp Cys Lys Leu Ile Leu Thr Leu Val Lys  
 20 25 30  
 Thr Gly Gly Leu Val Thr Ala Phe Val Tyr Val Ile Gly Val Ser Asn  
 35 40 45  
 Asp Phe Asn Met Leu Thr Thr His Lys Asn Ile Asn Phe Thr Ala Glu  
 50 55 60  
 Leu Phe Phe Asp Ser Thr Gly Asn Leu Leu Thr Ser Leu Ser Ser Leu  
 65 70 75 80  
 Lys Thr Pro Leu Asn His Lys Ser Gly Gln Asn Met Ala Thr Gly Ala  
 85 90 95  
 Leu Thr Asn Ala Lys Gly Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe  
 100 105 110  
 Asn Val Asn Ser Arg Glu Lys Glu Asn Tyr Ile Tyr Gly Thr Cys Tyr  
 115 120 125  
 Tyr Thr Ala Ser Asp His Thr Ala Phe Pro Ile Asp Ile Ser Val Met  
 130 135 140



16

Leu Asn Gln Arg Ala Leu Asn Asn Glu Thr Ser Tyr Cys Ile Arg Val  
 145 150 155 160  
 Thr Trp Ser Trp Asn Thr Gly Val Ala Pro Glu Val Gln Thr Ser Ala  
 165 170 175  
 Thr Thr Leu Val Thr Ser Pro Phe Thr Phe Tyr Tyr Ile Arg Glu Asp  
 180 185 190

Asp

<210> 21  
 <211> 193  
 <212> PRT  
 <213> Human adenovirus serotype 11A

&lt;400&gt; 21

Thr Leu Trp Thr Gly Ile Asn Pro Thr Glu Ala Asn Cys Gln Met Met  
 1 5 10 15  
 Asp Ser Ser Glu Ser Asn Asp Cys Lys Leu Ile Leu Thr Leu Val Lys  
 20 25 30  
 Thr Gly Ala Leu Val Thr Ala Phe Val Tyr Val Ile Gly Val Ser Asn  
 35 40 45  
 Asn Phe Asn Met Leu Thr Thr Tyr Arg Asn Ile Asn Phe Thr Ala Glu  
 50 55 60  
 Leu Phe Phe Asp Ser Ala Gly Asn Leu Leu Thr Ser Leu Ser Ser Leu  
 65 70 75 80  
 Lys Thr Pro Leu Asn His Lys Ser Gly Gln Asn Met Ala Thr Gly Ala  
 85 90 95  
 Ile Thr Asn Ala Lys Ser Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe  
 100 105 110  
 Asn Asn Asn Ser Arg Glu Lys Glu Asn Tyr Ile Tyr Gly Thr Cys His  
 115 120 125  
 Tyr Thr Ala Ser Asp His Thr Ala Phe Pro Ile Asp Ile Ser Val Met  
 130 135 140  
 Leu Asn Gln Arg Ala Ile Arg Ala Asp Thr Ser Tyr Cys Ile Arg Ile  
 145 150 155 160  
 Thr Trp Ser Trp Asn Thr Gly Asp Ala Pro Glu Gly Gln Thr Ser Ala  
 165 170 175  
 Thr Thr Leu Val Thr Ser Pro Phe Thr Phe Tyr Tyr Ile Arg Glu Asp  
 180 185 190

Asp

<210> 22  
 <211> 192  
 <212> PRT  
 <213> Human adenovirus serotype 16

&lt;400&gt; 22

Thr Leu Trp Thr Gly Ala Lys Pro Ser Ala Asn Cys Val Ile Lys Glu  
 1 5 10 15  
 Gly Glu Asp Ser Pro Asp Cys Lys Leu Thr Leu Val Leu Val Lys Asn  
 20 25 30  
 Gly Gly Leu Ile Asn Gly Tyr Ile Thr Leu Met Gly Ala Ser Glu Tyr  
 35 40 45

17

Thr Asn Thr Leu Phe Lys Asn Asn Gln Val Thr Ile Asp Val Asn Leu  
 50 55 60  
 Ala Phe Asp Asn Thr Gly Gln Ile Ile Thr Tyr Leu Ser Ser Leu Lys  
 65 70 75 80  
 Ser Asn Leu Asn Phe Lys Asp Asn Gln Asn Met Ala Thr Gly Thr Ile  
 85 90 95  
 Thr Ser Ala Lys Gly Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe Ile  
 100 105 110  
 Thr Tyr Ala Thr Glu Thr Leu Asn Glu Asp Tyr Ile Tyr Gly Glu Cys  
 115 120 125  
 Tyr Tyr Lys Ser Thr Asn Gly Thr Leu Phe Pro Leu Lys Val Thr Val  
 130 135 140  
 Thr Leu Asn Arg Arg Met Leu Ala Ser Gly Met Ala Tyr Ala Met Asn  
 145 150 155 160  
 Phe Ser Trp Ser Leu Asn Ala Glu Glu Ala Pro Glu Thr Thr Glu Val  
 165 170 175  
 Thr Leu Ile Thr Ser Pro Phe Phe Phe Ser Tyr Ile Arg Glu Asp Asp  
 180 185 190

<210> 23  
 <211> 191  
 <212> PRT  
 <213> Human adenovirus serotype 21

<400> 23

Thr Leu Trp Thr Gly Ile Lys Pro Pro Pro Asn Cys Gln Ile Val Glu  
 1 5 10 15  
 Asn Thr Asp Thr Asn Asp Gly Lys Leu Thr Leu Val Leu Val Lys Asn  
 20 25 30  
 Gly Gly Leu Val Asn Gly Tyr Val Ser Leu Val Gly Val Ser Asp Thr  
 35 40 45  
 Val Asn Gln Met Phe Thr Gln Lys Ser Ala Thr Ile Gln Leu Arg Leu  
 50 55 60  
 Tyr Phe Asp Ser Ser Gly Asn Leu Leu Thr Asp Glu Ser Asn Leu Lys  
 65 70 75 80  
 Ile Pro Leu Lys Asn Lys Ser Ser Thr Ala Thr Ser Glu Val Leu Gln  
 85 90 95  
 Pro Ala Glu Ala Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe Asn Thr  
 100 105 110  
 Thr Thr Arg Asp Ser Glu Asn Tyr Ile His Gly Ile Cys Tyr Tyr Met  
 115 120 125  
 Thr Ser Tyr Asp Arg Ser Leu Val Pro Leu Asn Ile Ser Ile Met Leu  
 130 135 140  
 Asn Ser Arg Thr Ile Ser Ser Asn Val Ala Tyr Ala Ile Gln Phe Glu  
 145 150 155 160  
 Trp Asn Leu Asn Ala Lys Glu Ser Pro Glu Ser Asn Ile Ala Thr Leu  
 165 170 175  
 Thr Thr Ser Pro Phe Phe Phe Ser Tyr Ile Arg Glu Asp Asp Asn  
 180 185 190

18

<210> 24  
 <211> 193  
 <212> PRT  
 <213> Human adenovirus serotype 34A

<400> 24

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Thr Leu Trp Thr Gly Val Asn Pro Thr Glu Ala Asn Cys Gln Ile Met
1      5      10      15
Asn Ser Ser Glu Ser Asn Asp Cys Lys Leu Ile Leu Thr Leu Val Lys
20      25      30
Thr Gly Ala Leu Val Thr Ala Phe Val Tyr Val Ile Gly Val Ser Asn
35      40      45
Asn Phe Asn Met Leu Thr Thr His Arg Asn Ile Asn Phe Thr Ala Glu
50      55      60
Leu Phe Phe Asp Ser Thr Gly Asn Leu Leu Thr Arg Leu Ser Ser Leu
65      70      75      80
Lys Thr Pro Leu Asn His Lys Ser Gly Gln Asn Met Ala Thr Gly Ala
85      90      95
Ile Thr Asn Ala Lys Gly Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe
100     105     110
Asn Asp Asn Ser Arg Glu Lys Glu Asn Tyr Ile Tyr Gly Thr Cys Tyr
115     120     125
Tyr Thr Ala Ser Asp His Thr Ala Phe Pro Ile Asp Ile Ser Val Met
130     135     140
Leu Asn Arg Arg Ala Ile Asn Asp Glu Thr Ser Tyr Cys Ile Arg Ile
145     150     155     160
Thr Trp Ser Trp Asn Thr Gly Asp Ala Pro Glu Val Gln Thr Ser Ala
165     170     175
Thr Thr Leu Val Thr Ser Pro Phe Thr Phe Tyr Tyr Ile Arg Glu Asp
180     185     190

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Asp

<210> 25  
 <211> 191  
 <212> PRT  
 <213> Human adenovirus serotype 35

<400> 25

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Thr Leu Trp Thr Gly Ile Asn Pro Pro Pro Asn Cys Gln Ile Val Glu
1      5      10
Asn Thr Asn Thr Asn Asp Gly Lys Leu Thr Leu Val Leu Val Lys Asn
20      25      30
Gly Gly Leu Val Asn Gly Tyr Val Ser Leu Val Gly Val Ser Asp Thr
35      40      45
Val Asn Gln Met Phe Thr Gln Lys Thr Ala Asn Ile Gln Leu Arg Leu
50      55      60
Tyr Phe Asp Ser Ser Gly Asn Leu Leu Thr Glu Glu Ser Asp Leu Lys
65      70      75      80
Ile Pro Leu Lys Asn Lys Ser Ser Thr Ala Thr Ser Glu Thr Val Ala
85      90      95
Ser Ser Lys Ala Phe Met Pro Ser Thr Thr Ala Tyr Pro Phe Asn Thr

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19

100 105 110

Thr Thr Arg Asp Ser Glu Asn Tyr Ile His Gly Ile Cys Tyr Tyr Met  
115 120 125

Thr Ser Tyr Asp Arg Ser Leu Phe Pro Leu Asn Ile Ser Ile Met Leu  
130 135 140

Asn Ser Arg Met Ile Ser Ser Asn Val Ala Tyr Ala Ile Gln Phe Glu  
145 150 155 160

Trp Asn Leu Asn Ala Ser Glu Ser Pro Glu Ser Asn Ile Ala Thr Leu  
165 170 175

Thr Thr Ser Pro Phe Phe Phe Ser Tyr Ile Thr Glu Asp Asp Asn  
180 185 190

<210> 26  
 <211> 156  
 <212> PRT  
 <213> Human adenovirus serotype 40SHORT

<400> 26

Thr Ile Trp Ser Ile Ser Pro Thr Pro Asn Cys Ser Ile Tyr Glu Thr  
1 5 10 15

Gln Asp Ala Asn Leu Phe Leu Cys Leu Thr Lys Asn Gly Ala His Val  
20 25 30

Leu Gly Thr Ile Thr Ile Lys Gly Leu Lys Gly Ala Leu Arg Glu Met  
35 40 45

Asn Asp Asn Ala Leu Ser Val Lys Leu Pro Phe Asp Asn Gln Gly Asn  
50 55 60

Leu Leu Asn Cys Ala Leu Glu Ser Ser Thr Trp Arg Tyr Gln Glu Thr  
65 70 75 80

Asn Ala Val Ala Ser Asn Ala Leu Thr Phe Met Pro Asn Ser Thr Val  
85 90 95

Tyr Pro Arg Asn Lys Thr Ala Asp Pro Gly Asn Met Leu Ile Gln Ile  
100 105 110

Ser Pro Asn Ile Thr Phe Ser Val Val Tyr Asn Glu Ile Asn Ser Gly  
115 120 125

Tyr Ala Phe Thr Phe Lys Trp Ser Ala Glu Pro Gly Lys Pro Phe His  
130 135 140

Pro Pro Thr Ala Val Phe Cys Tyr Ile Thr Glu Gln  
145 150 155

<210> 27  
 <211> 156  
 <212> PRT  
 <213> Human adenovirus serotype 41SHORT

<400> 27

Thr Ile Trp Ser Ile Ser Pro Thr Pro Asn Cys Ser Ile Tyr Glu Thr  
1 5 10 15

Gln Asp Ala Asn Leu Phe Leu Cys Leu Thr Lys Asn Gly Ala His Val  
20 25 30

Leu Gly Thr Ile Thr Ile Lys Gly Leu Lys Gly Ala Leu Arg Glu Met  
35 40 45

His Asp Asn Ala Leu Ser Leu Lys Leu Pro Phe Asp Asn Gln Gly Asn

20

50                                      55                                      60  
 Leu Leu Asn Cys Ala Leu Glu Ser Ser Thr Trp Arg Tyr Gln Glu Thr  
 65                                      70                                      75                                      80  
 Asn Ala Val Ala Ser Asn Ala Leu Thr Phe Met Pro Asn Ser Thr Val  
                                     85                                      90                                      95  
 Tyr Pro Arg Asn Lys Thr Ala His Pro Gly Asn Met Leu Ile Gln Ile  
                                     100                                      105                                      110  
 Ser Pro Asn Ile Thr Phe Ser Val Val Tyr Asn Glu Ile Asn Ser Gly  
                                     115                                      120                                      125  
 Tyr Ala Phe Thr Phe Lys Trp Ser Ala Glu Pro Gly Lys Pro Phe His  
                                     130                                      135                                      140  
 Pro Pro Thr Ala Val Phe Cys Tyr Ile Thr Glu Gln  
 145                                      150                                      155  
  
 <210> 28  
 <211> 354  
 <212> PRT  
 <213> Anti-HA ScFv fused in frame with 2 C-terminal myc epitopes and  
                                     PDGF receptor transmembrane anchor (Anti-HA pseudo-receptor)  
  
 <400> 28  
 Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro  
   1                                      5                                      10                                      15  
 Gly Ser Thr Gly Asp Gly Ala Gln Pro Ala Asp Ile Val Met Thr Gln  
                                     20                                      25                                      30  
 Ser Pro Ser Ser Leu Thr Val Thr Ala Gly Glu Lys Val Thr Met Ser  
                                     35                                      40                                      45  
 Cys Lys Ser Ser Gln Ser Leu Leu Asn Ser Gly Asn Gln Lys Asn Tyr  
                                     50                                      55                                      60  
 Leu Thr Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile  
   65                                      70                                      75                                      80  
 Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Gly  
                                     85                                      90                                      95  
 Ser Gly Ser Gly Arg Asp Phe Thr Leu Thr Ile Ser Ser Val Gln Ala  
                                     100                                      105                                      110  
 Glu Asp Leu Ala Val Tyr Tyr Cys Gln Asn Asp Asn Ser His Pro Leu  
                                     115                                      120                                      125  
 Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu Lys Arg Ala Gly Gly Gly  
                                     130                                      135                                      140  
 Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Glu Val Gln Leu  
   145                                      150                                      155                                      160  
 Val Glu Ser Gly Gly Asn Leu Val Asn Pro Gly Gly Ser Leu Lys Leu  
                                     165                                      170                                      175  
 Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Thr Tyr Gly Met Ser Trp  
                                     180                                      185                                      190  
 Val Arg Gln Thr Pro Asn Lys Arg Leu Glu Trp Val Pro Thr Ile Ile  
                                     195                                      200                                      205  
 Arg Gly Gly Ser Tyr Thr Tyr Tyr Pro Asp Ser Val Lys Gly Arg Phe  
                                     210                                      215                                      220  
 Thr Ile Ser Lys Asn Asn Ala Lys Asn Thr Leu Tyr Leu Gln Met Ser  
   225                                      230                                      235                                      240

21

Ser Leu Lys Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Lys Arg Glu  
 245 250 255  
 Thr Phe Asp Glu Lys Gly Phe Ala Tyr Trp Gly Gln Gly Thr Leu Val  
 260 265 270  
 Thr Val Ser Ala Ala Ala Ala Glu Gln Lys Leu Ile Ser Glu Glu Asp  
 275 280 285  
 Leu Asn Gly Ala Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu  
 290 295 300  
 Asn Ala Val Gly Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser  
 305 310 315 320  
 Leu Pro Phe Lys Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val  
 325 330 335  
 Leu Thr Ile Ile Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys  
 340 345 350

Pro Val.

&lt;210&gt; 29

&lt;211&gt; 218

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: Consensus  
 sequence from the comparison between non-group B  
 adenoviral knobs as indicated in Figures 1A and  
 1B. Xaa is any amino acid or no amino acid as  
 indicated in Figures 1A and 1B.

&lt;400&gt; 29

Thr Leu Trp Thr Thr Pro Xaa Pro Ser Pro Asn Cys Xaa Xaa Xaa Xaa  
 1 5 10 15  
 Xaa Lys Asp Xaa Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30  
 Ile Leu Ala Xaa Val Ser Xaa Xaa Xaa Val Xaa Xaa Xaa Xaa Gly Xaa  
 35 40 45  
 Xaa Xaa Xaa Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa  
 50 55 60  
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa  
 65 70 75 80  
 Xaa Xaa Xaa Xaa Phe Asp Xaa Asn Gly Val Leu Xaa Xaa Xaa Ser Xaa  
 85 90 95  
 Xaa Xaa Xaa Leu Xaa Xaa Xaa Tyr Trp Asn Phe Arg Xaa Gly Xaa Xaa  
 100 105 110  
 Xaa Xaa Xaa Xaa Xaa Tyr Xaa Asn Ala Val Gly Phe Met Pro Asn Xaa  
 115 120 125  
 Xaa Ala Tyr Pro Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa  
 130 135 140  
 Ala Xaa Xaa Xaa Xaa Ile Val Xaa Xaa Xaa Xaa Tyr Leu Xaa Gly Xaa  
 145 150 155 160  
 Xaa Xaa Xaa Pro Xaa Xaa Xaa Xaa Xaa Thr Xaa Asn Xaa Xaa Xaa Glu  
 165 170 175

22

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Tyr Ser Xaa Xaa Phe Xaa Xaa  
 180 185 190

Xaa Trp Xaa Xaa Xaa Xaa Xaa Tyr Xaa Asn Xaa Xaa Phe Xaa Thr Xaa  
 195 200 205

Ser Xaa Thr Phe Ser Tyr Ile Ala Gln Glu  
 210 215

&lt;210&gt; 30

&lt;211&gt; 215

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: Majority  
 sequence from the comparison between non-group B  
 adenoviral knobs as indicated in Figures 1A and  
 1B. Xaa is any amino acid or no amino acid as  
 indicated in Figures 1A and 1B.

&lt;400&gt; 30

Thr Leu Trp Thr Thr Pro Asp Pro Ser Pro Asn Cys Thr Ile Asp Gln  
 1 5 10 15

Asp Lys Asp Ser Lys Leu Thr Leu Val Leu Thr Lys Cys Gly Ser Gln  
 20 25 30

Ile Leu Ala Thr Val Ser Leu Ile Val Val Xaa Ala Xaa Xaa Gly Lys  
 35 40 45

Leu Leu Ile Ile Asn Asn Thr Thr Asn Pro Xaa Xaa Xaa Xaa Xaa Xaa  
 50 55 60

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Lys Xaa Phe Thr  
 65 70 75 80

Ile Lys Leu Leu Phe Asp Ala Asn Gly Val Leu Leu Glu Asn Ser Asn  
 85 90 95

Xaa Xaa Xaa Leu Gly Lys Ala Tyr Trp Asn Phe Arg Asn Gly Asn Ser  
 100 105 110

Thr Val Ser Thr Ala Tyr Glu Asn Ala Val Gly Phe Met Pro Asn Leu  
 115 120 125

Val Ala Tyr Pro Lys Pro Thr Gly Xaa Ser Xaa Xaa Xaa Xaa Xaa Xaa  
 130 135 140

Ala Lys Asp Xaa Xaa Ile Val Tyr Gly Asn Val Tyr Leu Gly Gly Asp  
 145 150 155 160

Pro Asp Gln Pro Val Val Ile Lys Ile Thr Phe Asn Xaa Xaa Gln Glu  
 165 170 175

Thr Xaa Xaa Gly Ser Gly Tyr Ser Ile Thr Phe Asp Phe Ser Trp Ser  
 180 185 190

Lys Xaa Xaa Thr Tyr Ile Asn Val Glu Phe Glu Thr Thr Ser Phe Thr  
 195 200 205

Phe Ser Tyr Ile Ala Gln Glu  
 210 215

&lt;210&gt; 31

&lt;211&gt; 248

&lt;212&gt; PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Consensus sequence from the comparison between non-group B adenoviral knobs as indicated in Figures 2A and 2B. Xaa is any amino acid or no amino acid as indicated in Figures 2A and 2B.

<400> 31

```

Thr Leu Trp Thr Gly Xaa Asn Pro Xaa Xaa Ala Asn Cys Gln Ile Xaa
 1          5          10          15
Xaa Xaa Xaa Xaa Ser Asn Asp Cys Lys Leu Thr Leu Xaa Leu Val Lys
          20          25          30
Asn Gly Gly Leu Val Asn Gly Tyr Val Xaa Leu Xaa Gly Val Xaa Ser
          35          40          45
Xaa Xaa Xaa Asn Xaa Leu Xaa Xaa Phe Thr Xaa Lys Asn Xaa Xaa Xaa
          50          55          60
Xaa Xaa Xaa Asn Ile Xaa Xaa Glu Leu Xaa Xaa Xaa Xaa Xaa Xaa
          65          70          75          80
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Phe Asp Ser Thr Gly Asn
          85          90          95
Leu Leu Thr Xaa Xaa Xaa Xaa Xaa Leu Ser Ser Leu Xaa Xaa Xaa Lys
          100          105          110
Thr Pro Leu Asn Xaa Lys Ser Xaa Gln Asn Met Ala Thr Gly Ala Xaa
          115          120          125
Thr Xaa Ala Lys Gly Phe Met Pro Ser Thr Thr Ala Tyr Xaa Xaa Pro
          130          135          140
Phe Asn Xaa Xaa Xaa Arg Glu Xaa Xaa Xaa Xaa Xaa Xaa Glu Asn
          145          150          155          160
Tyr Ile Tyr Gly Xaa Cys Tyr Tyr Xaa Ala Ser Xaa Asp Xaa Thr Leu
          165          170          175
Phe Pro Leu Xaa Ile Ser Val Met Leu Asn Xaa Xaa Xaa Arg Xaa Ile
          180          185          190
Xaa Ser Xaa Xaa Xaa Xaa Xaa Xaa Xaa Thr Ser Xaa Xaa Tyr Xaa Ile
          195          200          205
Xaa Phe Xaa Trp Ser Leu Asn Ala Xaa Gly Xaa Ala Pro Xaa Xaa Glu
          210          215          220
Thr Xaa Xaa Xaa Xaa Xaa Xaa Thr Leu Xaa Thr Ser Pro Phe Thr Phe
          225          230          235          240
Ser Tyr Ile Arg Glu Asp Xaa Asp
          245

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<210> 32

<211> 248

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Majority sequence from the comparison between non-group B adenoviral knobs as indicated in Figures 2A and 2B. Xaa is any amino acid or no amino acid as



indicated in Figures 2A and 2B.

<400> 32  
 Thr Leu Trp Thr Gly Ile Asn Pro Xaa Glu Ala Asn Cys Gln Ile Met  
 1 5 10 15  
 Glu Ser Ser Glu Ser Asn Asp Cys Lys Leu Thr Leu Val Leu Val Lys  
 20 25 30  
 Asn Gly Gly Leu Val Asn Gly Tyr Val Tyr Leu Ile Gly Val Xaa Ser  
 35 40 45  
 Asp Thr Val Asn Met Leu Xaa Xaa Phe Thr Asn Lys Asn Xaa Xaa Xaa  
 50 55 60  
 Xaa Xaa Ile Asn Ile Thr Ala Glu Leu Xaa Xaa Xaa Xaa Xaa Xaa  
 65 70 75 80  
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Phe Phe Asp Ser Thr Gly Asn  
 85 90 95  
 Leu Leu Thr Xaa Xaa Xaa Xaa Ser Leu Ser Ser Leu Xaa Xaa Xaa Lys  
 100 105 110  
 Thr Pro Leu Asn His Lys Ser Gly Gln Asn Met Ala Thr Gly Ala Ile  
 115 120 125  
 Thr Ser Ala Lys Gly Phe Met Pro Ser Thr Thr Ala Tyr Xaa Xaa Pro  
 130 135 140  
 Phe Asn Thr Asn Ser Arg Glu Xaa Xaa Xaa Xaa Xaa Xaa Lys Glu Asn  
 145 150 155 160  
 Tyr Ile Tyr Gly Thr Cys Tyr Tyr Thr Ala Ser Xaa Asp His Thr Leu  
 165 170 175  
 Phe Pro Leu Asp Ile Ser Val Met Leu Asn Xaa Xaa Ser Arg Ala Ile  
 180 185 190  
 Ser Ser Glu Xaa Xaa Xaa Xaa Xaa Xaa Thr Ser Xaa Xaa Tyr Ala Ile  
 195 200 205  
 Arg Phe Thr Trp Ser Leu Asn Ala Xaa Gly Glu Ala Pro Xaa Xaa Glu  
 210 215 220  
 Thr Ser Xaa Xaa Xaa Ala Ala Thr Leu Val Thr Ser Pro Phe Thr Phe  
 225 230 235 240  
 Ser Tyr Ile Arg Glu Asp Xaa Asp  
 245

# INTERNATIONAL SEARCH REPORT

Intern al Application No  
PCT/US 99/20728

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/861 C07K14/075 C12N7/00

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 96 26281 A (WICKHAM, TJ; GENVEC INC; CORNELL RES FOUNDATION INC (US)) 29 August 1996 (1996-08-29) example 2 ---	1,4,5,8, 9
P,X	WO 98 44121 A (TRANSGENE SA; CNRS) 8 October 1998 (1998-10-08) page 8, line 4 - line 9 ---	1
P,X	WO 98 54346 A (GENVEC, INC.) 3 December 1998 (1998-12-03) page 3, line 3 - line 13; examples 1-4 ---	1
A	WO 98 13499 A (CIBA GEIGY AG; SCRIPPS RESEARCH INST (US); MEMEROW GR (US); VO) 2 April 1998 (1998-04-02) page 59, paragraph 3 --- -/--	1,9-11

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

\* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

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"P" document published prior to the international filing date but later than the priority date claimed

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"&" document member of the same patent family

Date of the actual completion of the international search

10 January 2000

Date of mailing of the international search report

17/01/2000

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Authorized officer

Cupido, M

# INTERNATIONAL SEARCH REPORT

Inter.: 1st Application No  
PCT/US 99/20728

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>WO 97 06826 A (BOTH GW; COMMW SCIENT IND RES ORG (AU)) 27 February 1997 (1997-02-27)</p> <p>page 5, line 4; figures 3,4</p> <p style="text-align: center;">---</p>	<p>1,4,5, 9-11, 22-27, 33,34</p>
A	<p>WICKHAM T J ET AL: "INCREASED IN VITRO AND IN VIVO GENE TRANSFER BY ADENOVIRUS VECTORS CONTAINING CHIMERIC FIBER PROTEINS" JOURNAL OF VIROLOGY, US, THE AMERICAN SOCIETY FOR MICROBIOLOGY, vol. 71, no. 11, page 8221-8229 XP002911344 ISSN: 0022-538X page 8223, right-hand column -page 8224, left-hand column</p> <p style="text-align: center;">-----</p>	<p>11-33</p>

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/ 20728

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:  
Remark: Although claims 33 and 34 insofar they concern an in vivo method are directed to a method of treatment of the human or animal body, the search has been carried out and based on the alleged effects of the adenoviral vector.
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

### Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

information on patent family members

Inter: International Application No

PCT/US 99/20728

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9626281 A	29-08-1996	US 5770442 A AU 698254 B AU 4980496 A CA 2213343 A EP 0811069 A JP 11500315 T	23-06-1998 29-10-1998 11-09-1996 29-08-1996 10-12-1997 12-01-1999
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WO 9854346 A	03-12-1998	AU 7604998 A	30-12-1998
WO 9813499 A	02-04-1998	AU 4624197 A EP 0937150 A	17-04-1998 25-08-1999
WO 9706826 A	27-02-1997	AU 708870 B AU 6696696 A CA 2229631 A EP 0851769 A JP 11511139 T NZ 315295 A	12-08-1999 12-03-1997 27-02-1997 08-07-1998 28-09-1999 29-09-1999